

*Transitioning
Research in the
Physical
Sciences to
Planetary
Science*

Paola Grantham
Jet Propulsion Laboratory
California Institute of
Technology

Structure & Chemistry

Imaging laser-induced breakdown

Atomic force microscopy + IR

Electron-induced X-ray spectroscopy

Fluorescence & Raman

Volatiles & gases

Molecular absorption spectroscopy

Lab-on-a-chip systems

Capillary electrophoresis

Force-detected NMR



Emerging astrobiology instruments



One approach for life detection—look for structure & chemistry

Macroscopic ----- Microscopic ----- “nanoscopic”

Find the structures

Determine their chemistry

Elemental composition

Chirality

Complex organic molecules

Isotopic fractionation

μCE with MOD

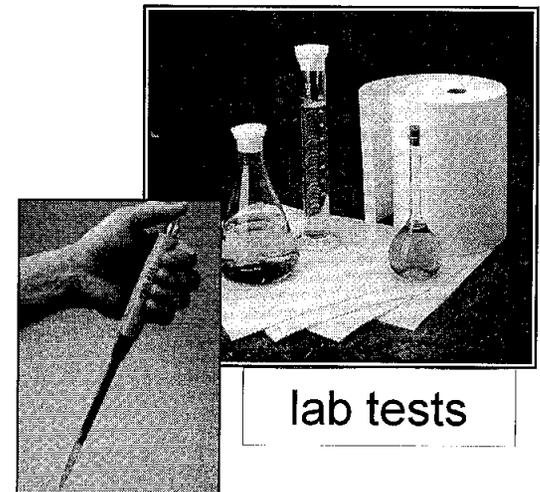
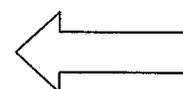
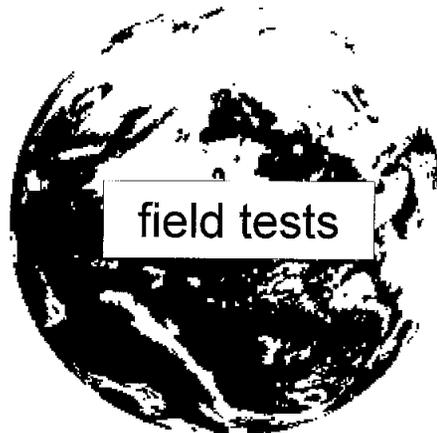
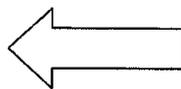
Imaging probe with LIBS

AFM with IR spectroscopies

UV Raman + fluorescence

FD-NMR

TDL absorption spectroscopy

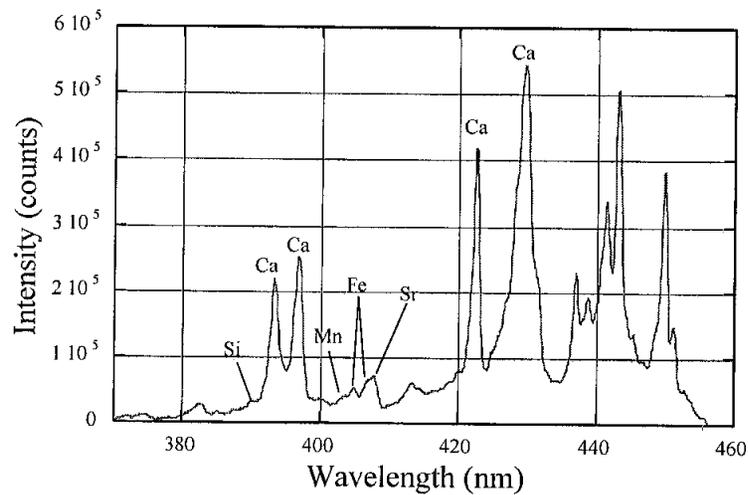
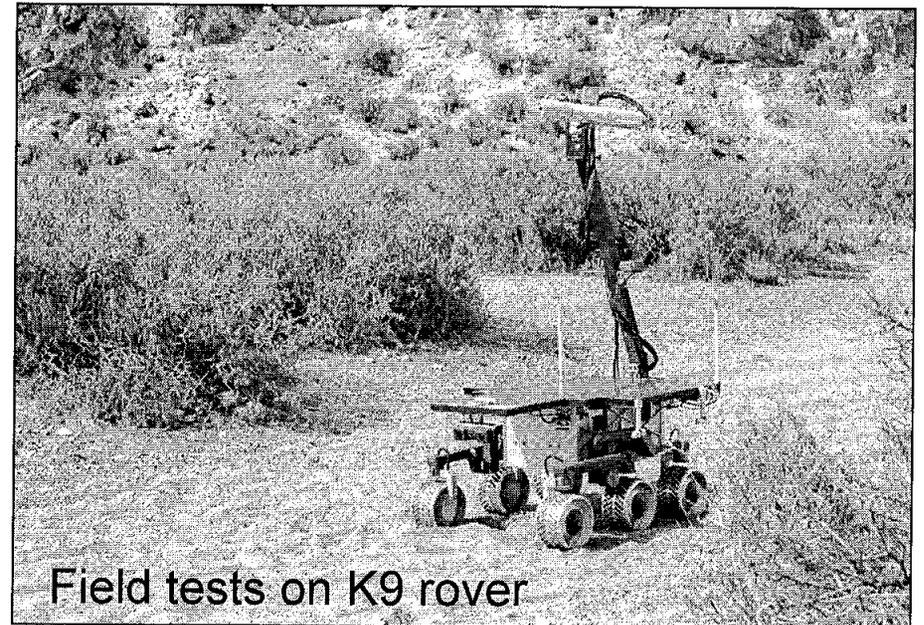
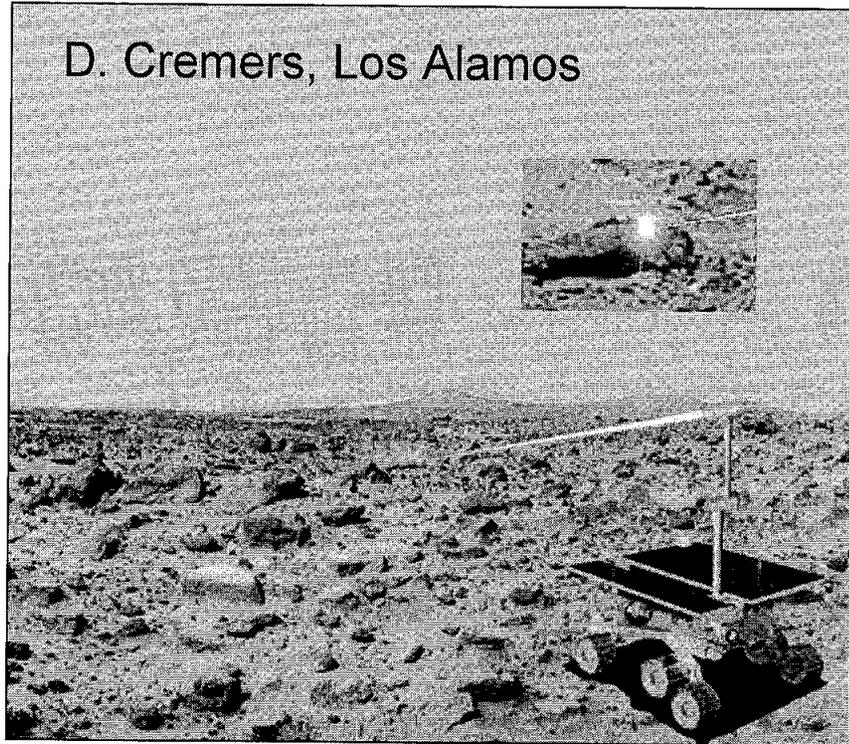
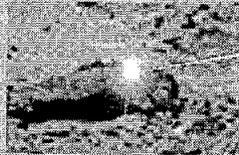




LIBS for "remote" analysis

JPL

D. Cremers, Los Alamos



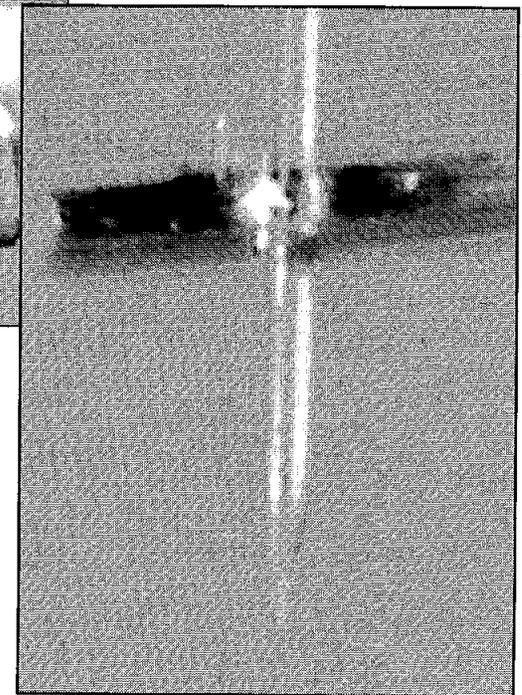
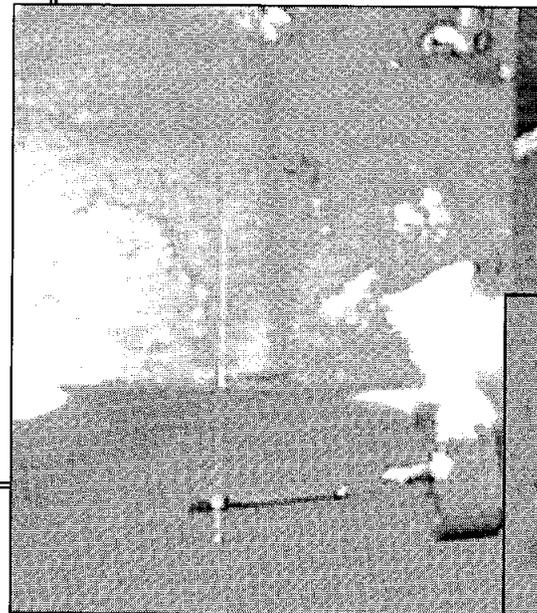


In-situ LIBS-based chemical imager

JPL

Dmitri Kossakovski
J. Kirschvink, Caltech

QuickTime™ and a
decompressor
are needed to see this picture.



- ◆ **Scanning probe microscopy coupled with laser-induced breakdown spectroscopy**
 - **Topographic imaging**
 - **Elemental analysis registered to the topography**



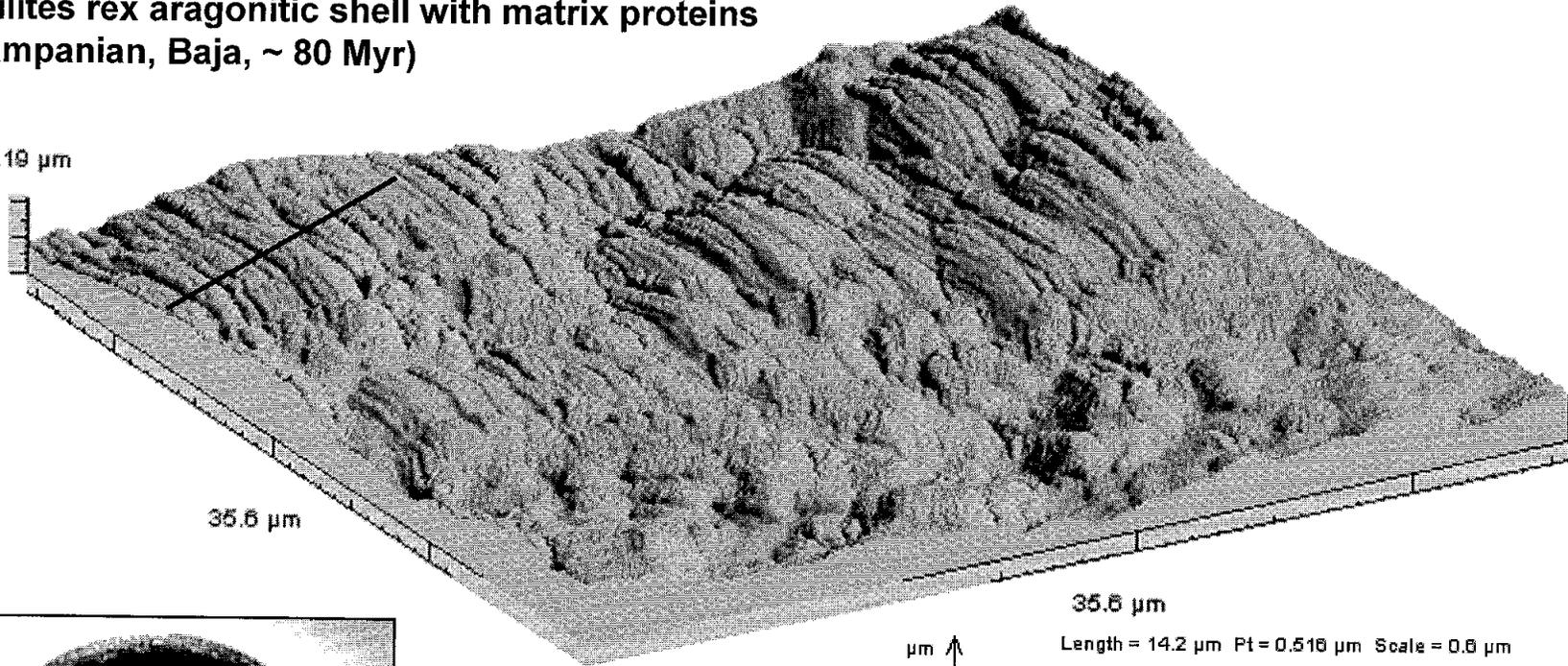
Topographic imaging

JPL

Alpha = 30° Beta = 20°

Bacculites rex aragonitic shell with matrix proteins
(U. Campanian, Baja, ~ 80 Myr)

2.19 μm

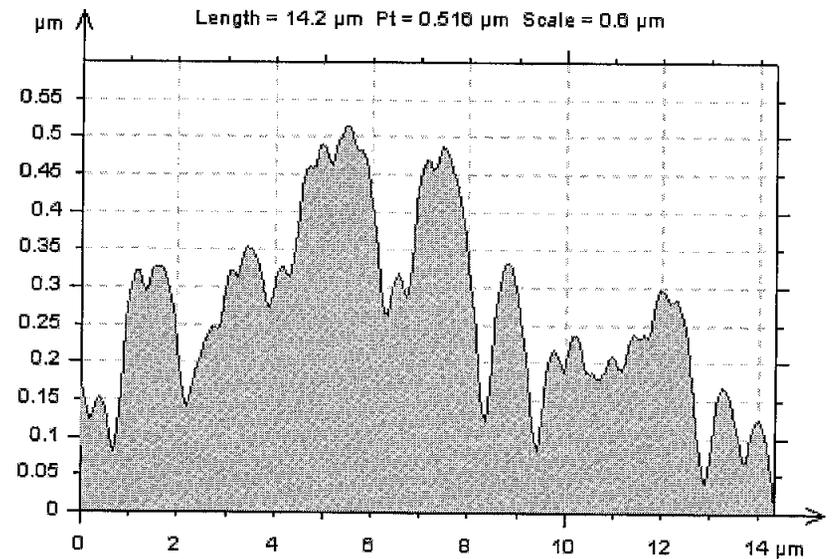


35.6 μm



35.6 μm

Length = 14.2 μm Pt = 0.516 μm Scale = 0.6 μm



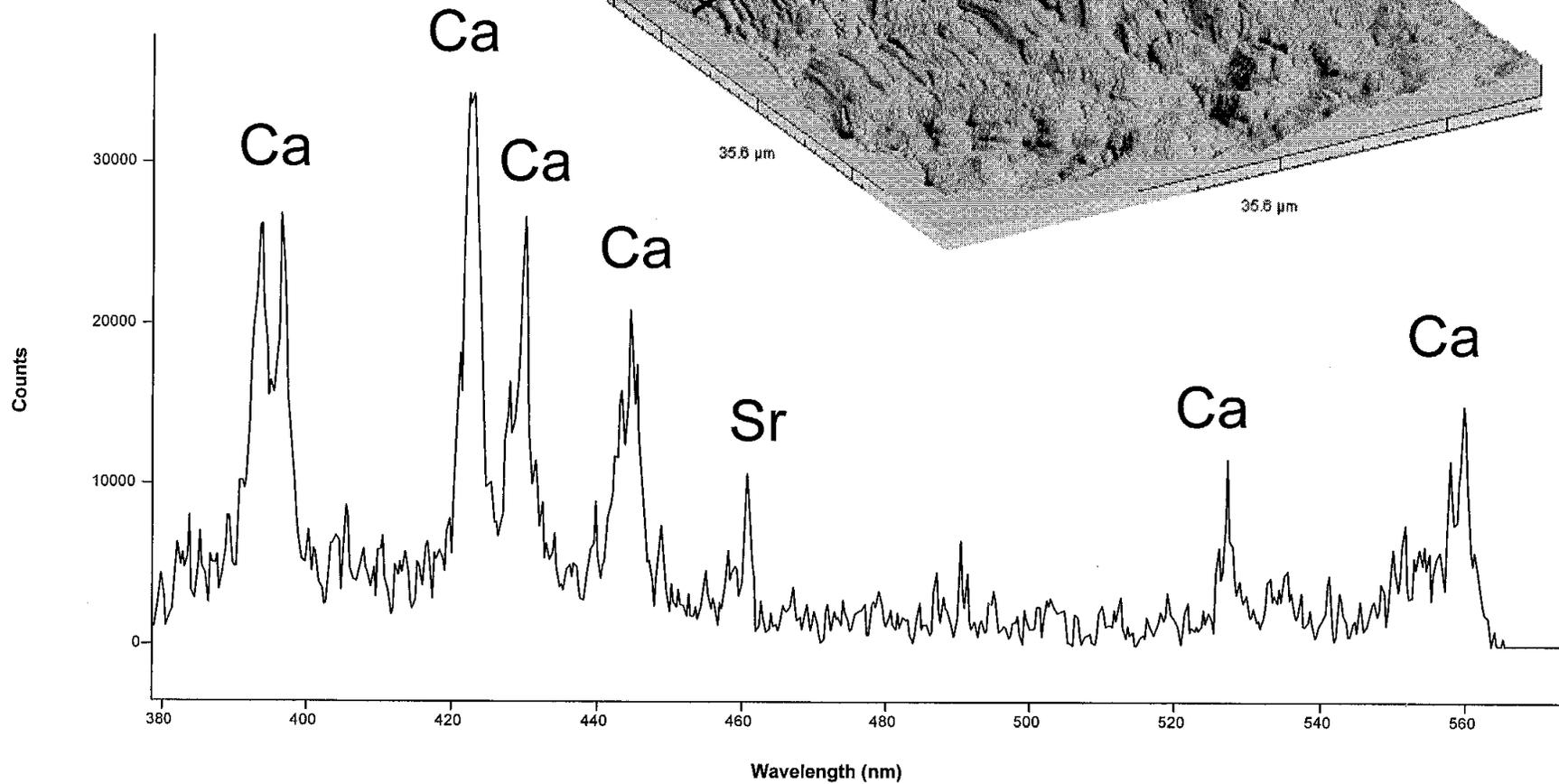
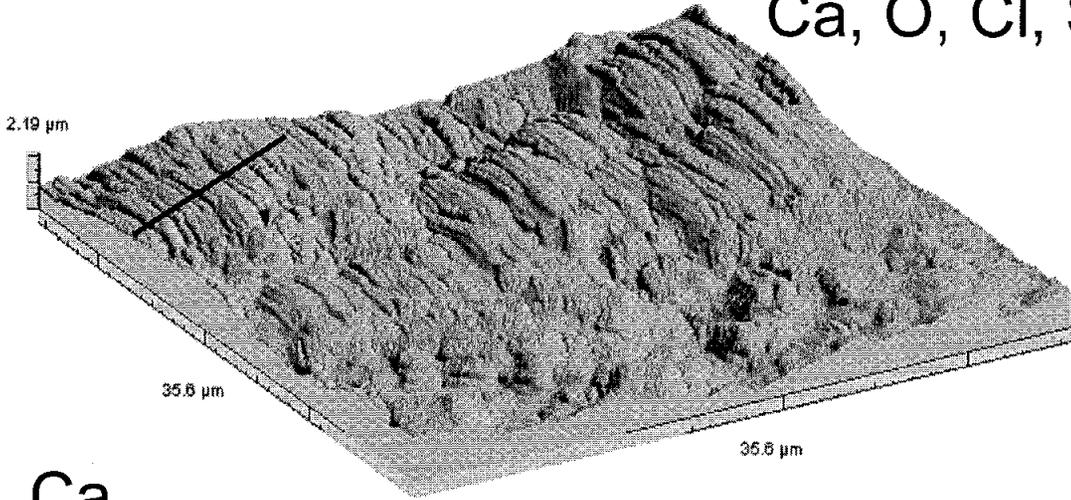


LIBS spectrum of fossil shell



SEM-EDS:
Ca, O, Cl, S, Sr

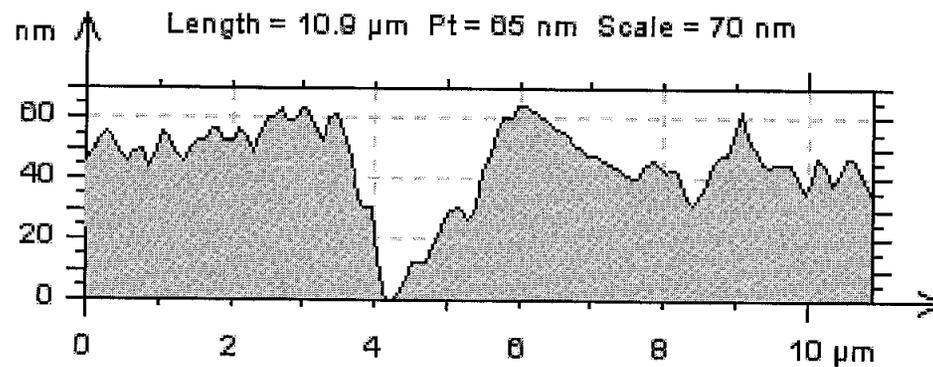
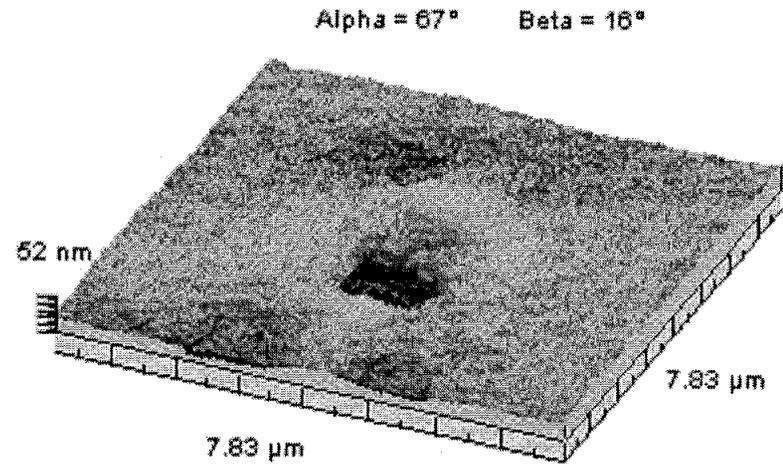
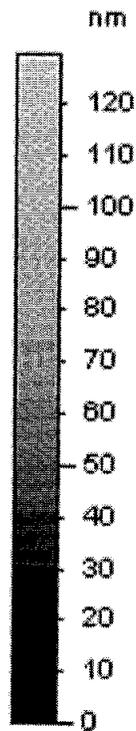
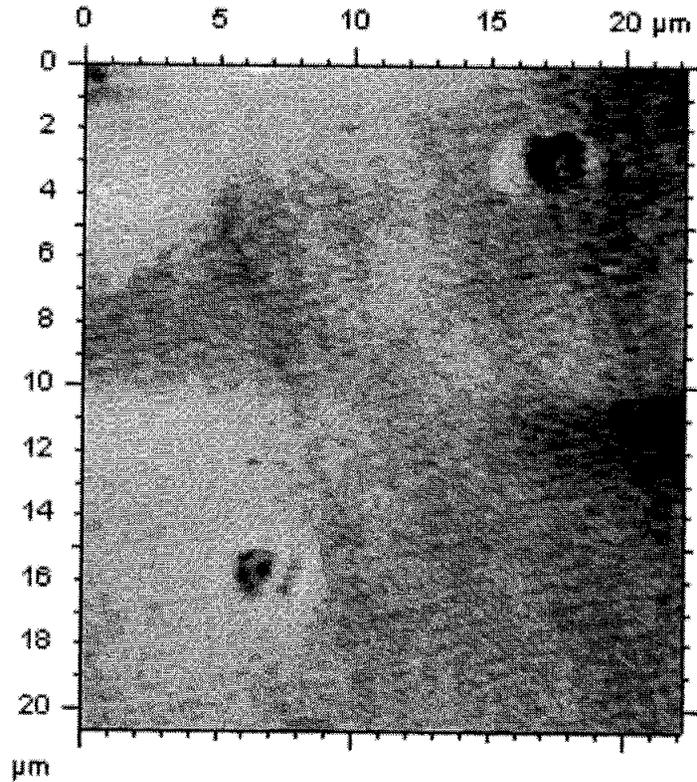
Alpha = 30° Beta = 20°





Crater Formation on polished Basalt

JPL

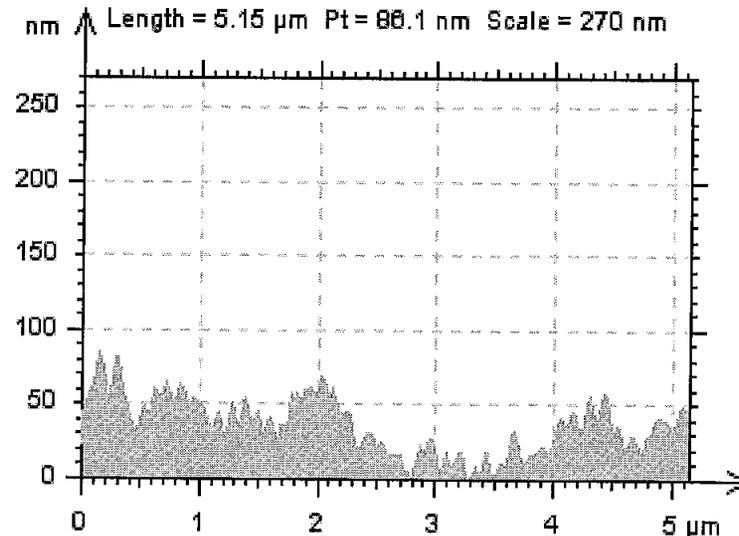
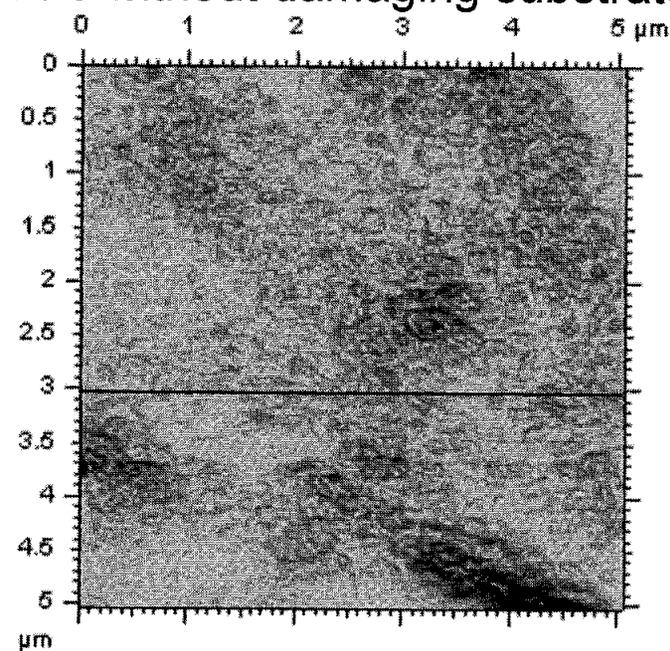
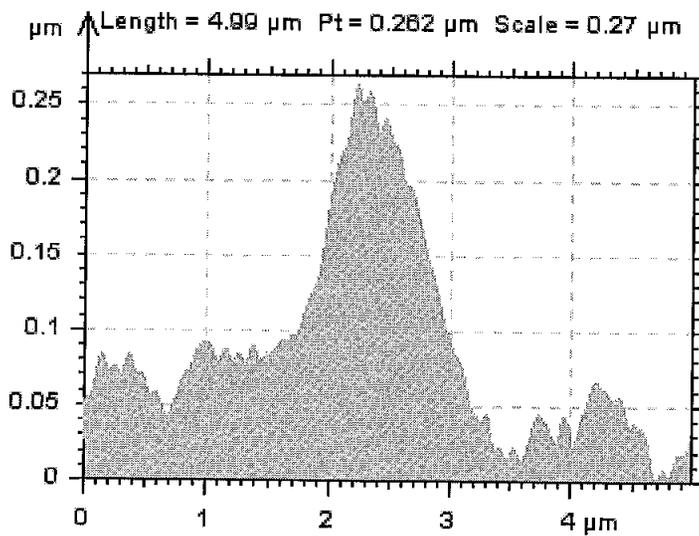
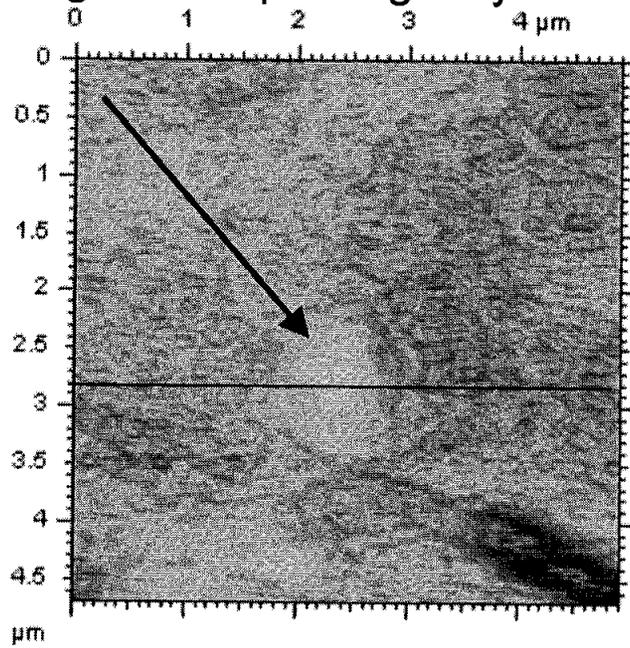




Particle study

JPL

Single laser pulse gently ablates particle without damaging substrate





B. subtilis spores on cleaned surface

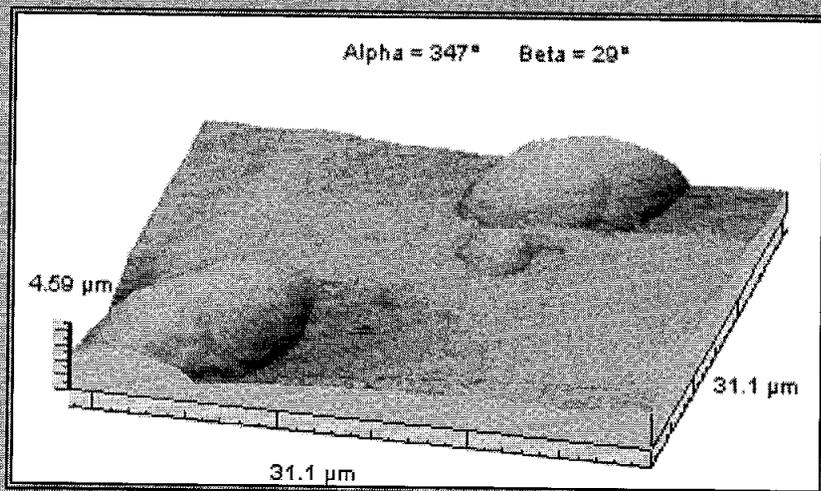


Image from JPL's Planetary Protection Group

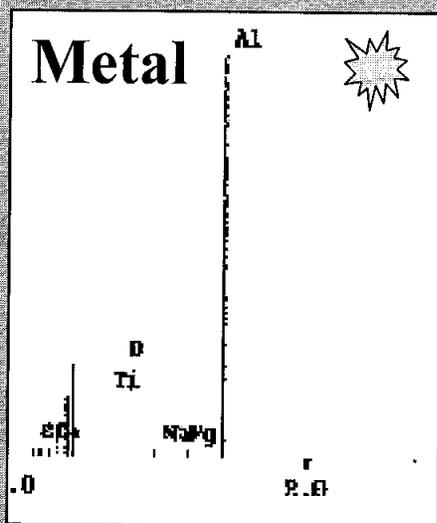
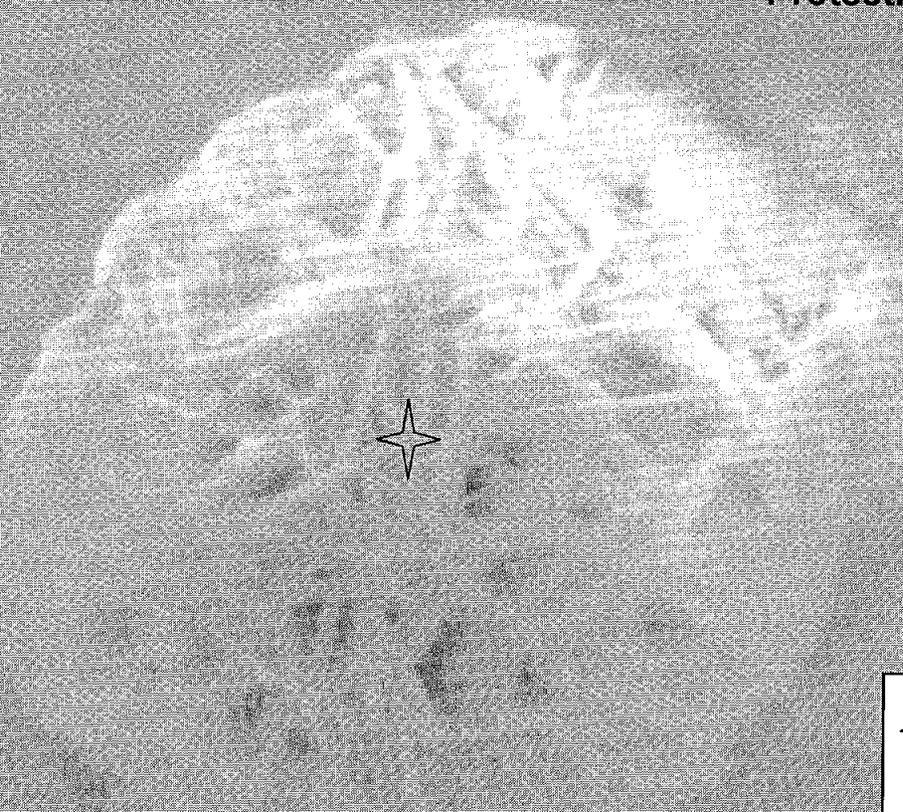
Acc V Spot Magn Det WD |-----| 20 μm
10.0 kV 3.0 1200x SE 11.5 Hiyac



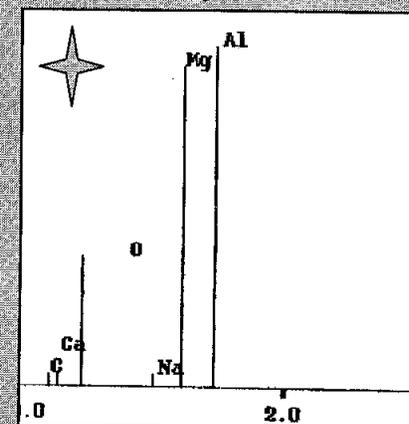
B. subtilis spores on Aluminum



Image from JPL's Planetary Protection Group



Crystal



Acc.V Spot Magn Det WD |-----| 2 μ m
5.00 kV 3.0 10000x SE 10.2 Hiyac

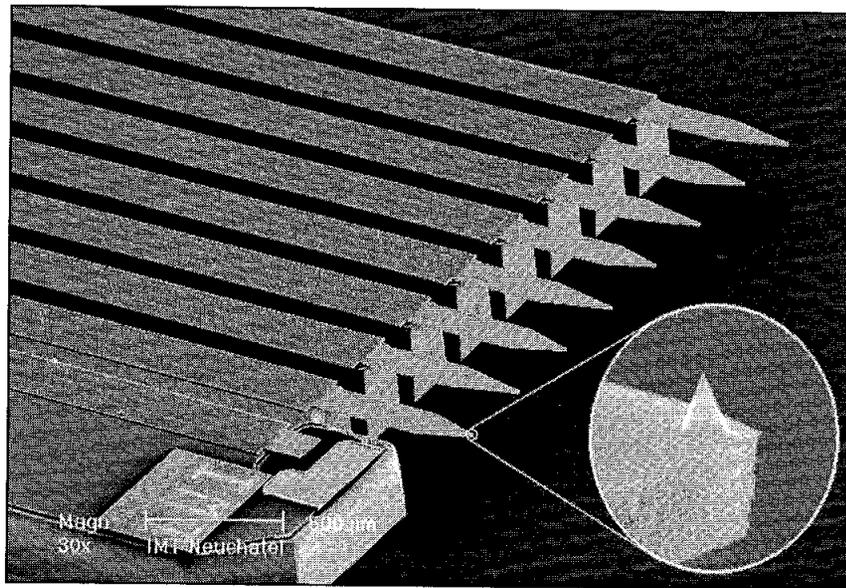
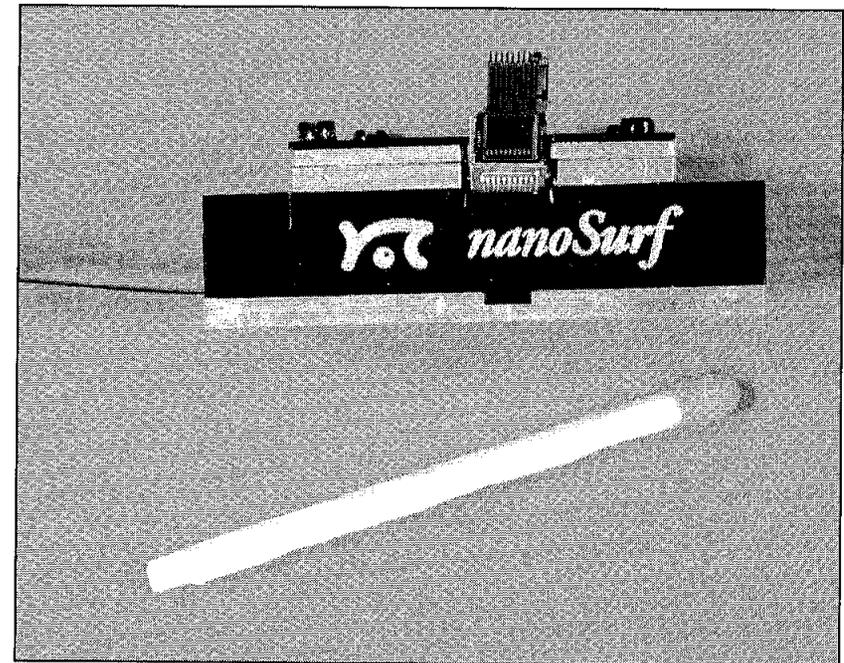
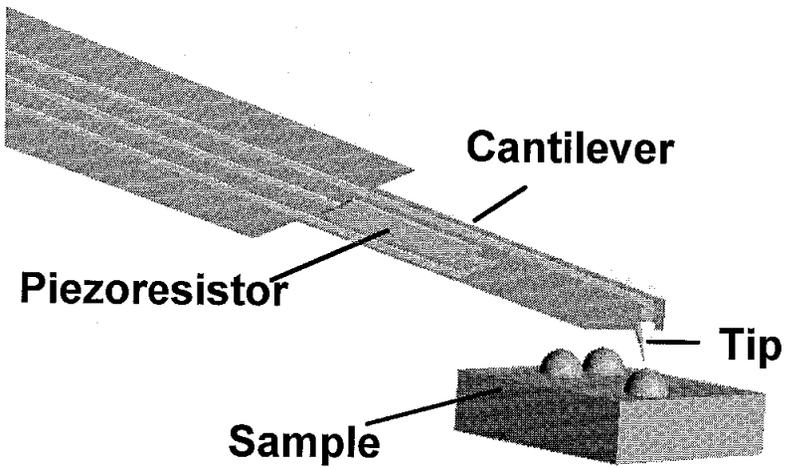


AFM for imaging & spectroscopy

JPL

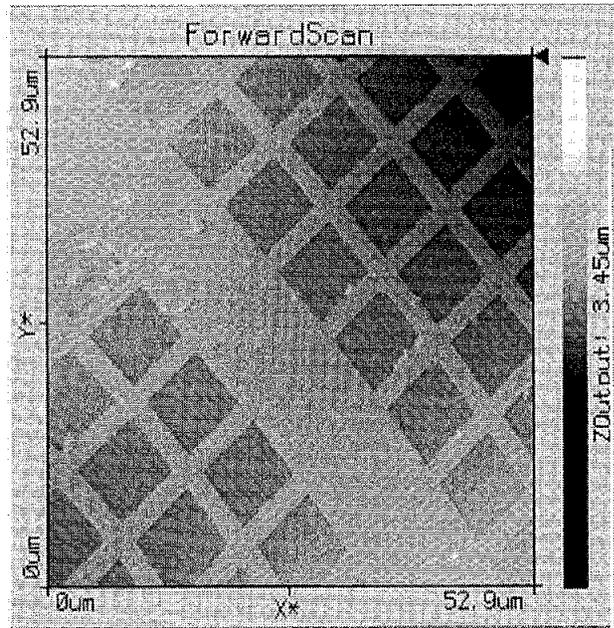
Sensor head

T. Pike & M. Anderson, JPL

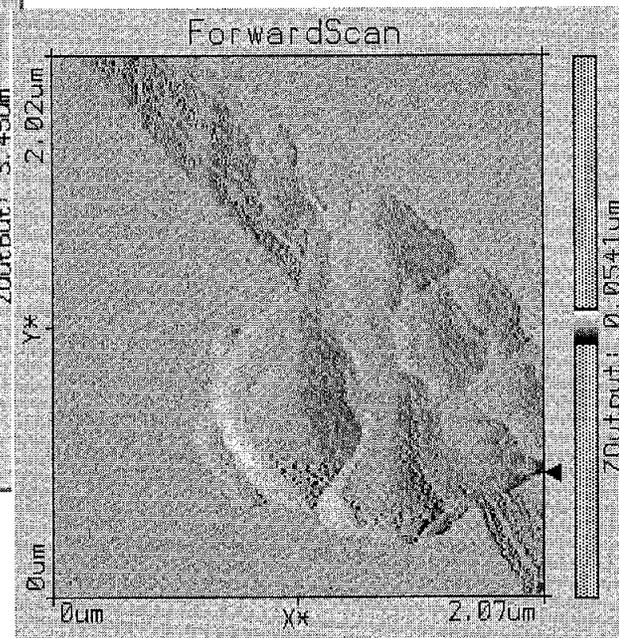




MECA AFM images



Large scan of calibration substrate with dust particles

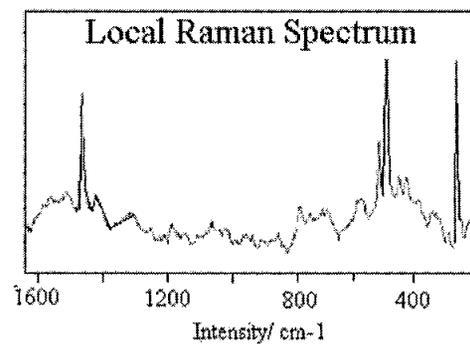
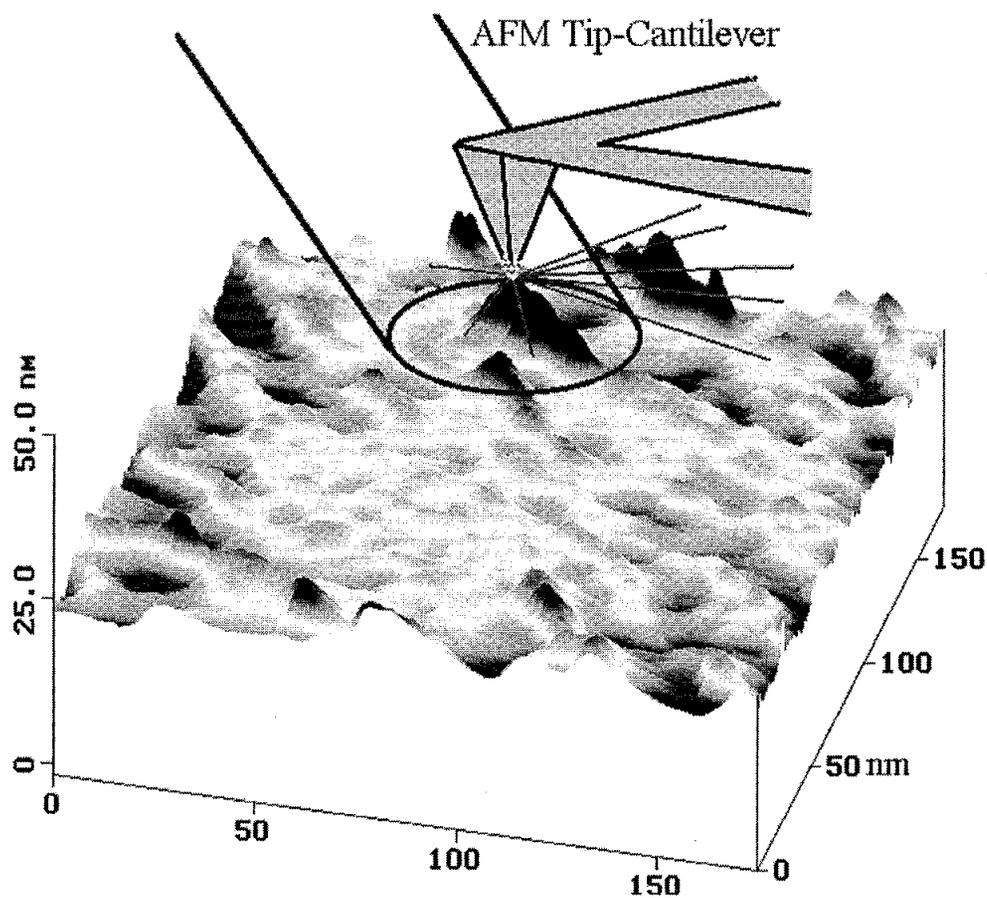


High-resolution scan of silicon etchant feature in calibration substrate

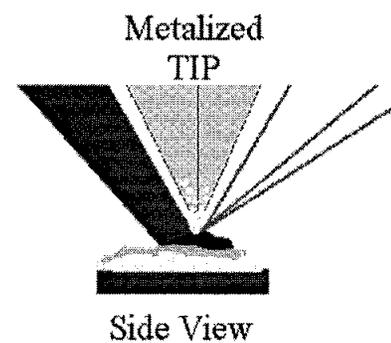
Scan of fragment of diatom



Surface-Enhanced Raman with AFM



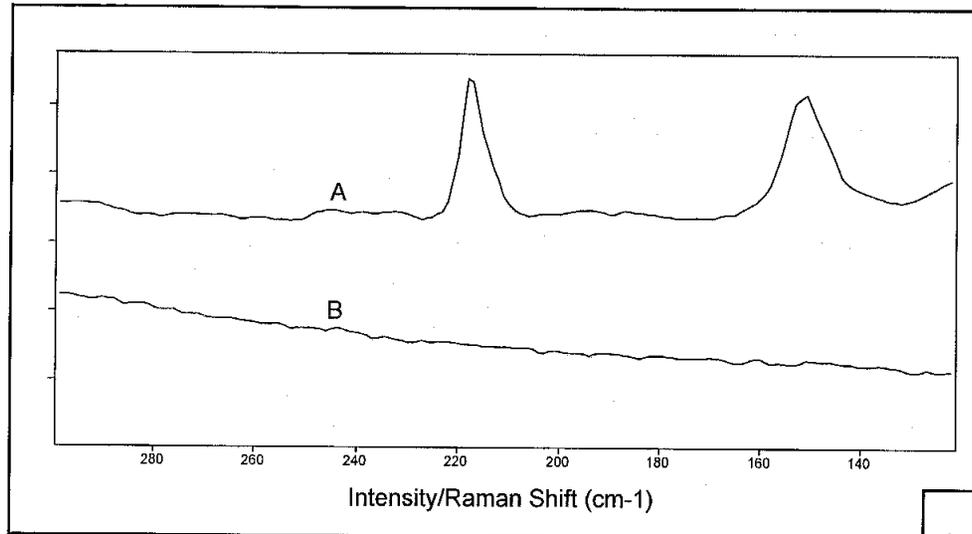
- Laser Excitation Beam
- Enhanced Raman Signal





AFM for surface-enhanced Raman

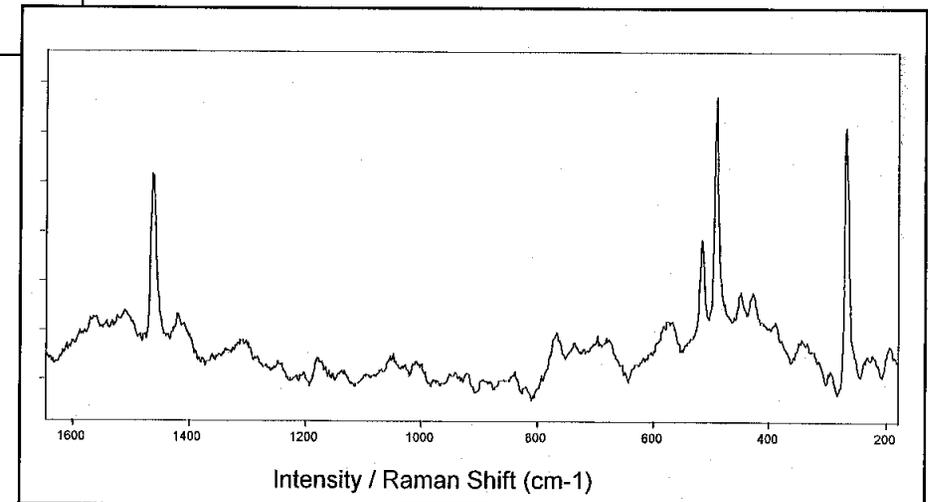
JPL



Gold-coated AFM tip generating a local surface enhanced Raman effect on a 10 μm sulfur film (A).

When the beam is focused away from the tip on the film, the Raman signal is undetectable using the same microprobe parameters (B).

Raman spectrum of ~10 femtograms residue from the surface of a C_{60} sample surgically removed with AFM tip.



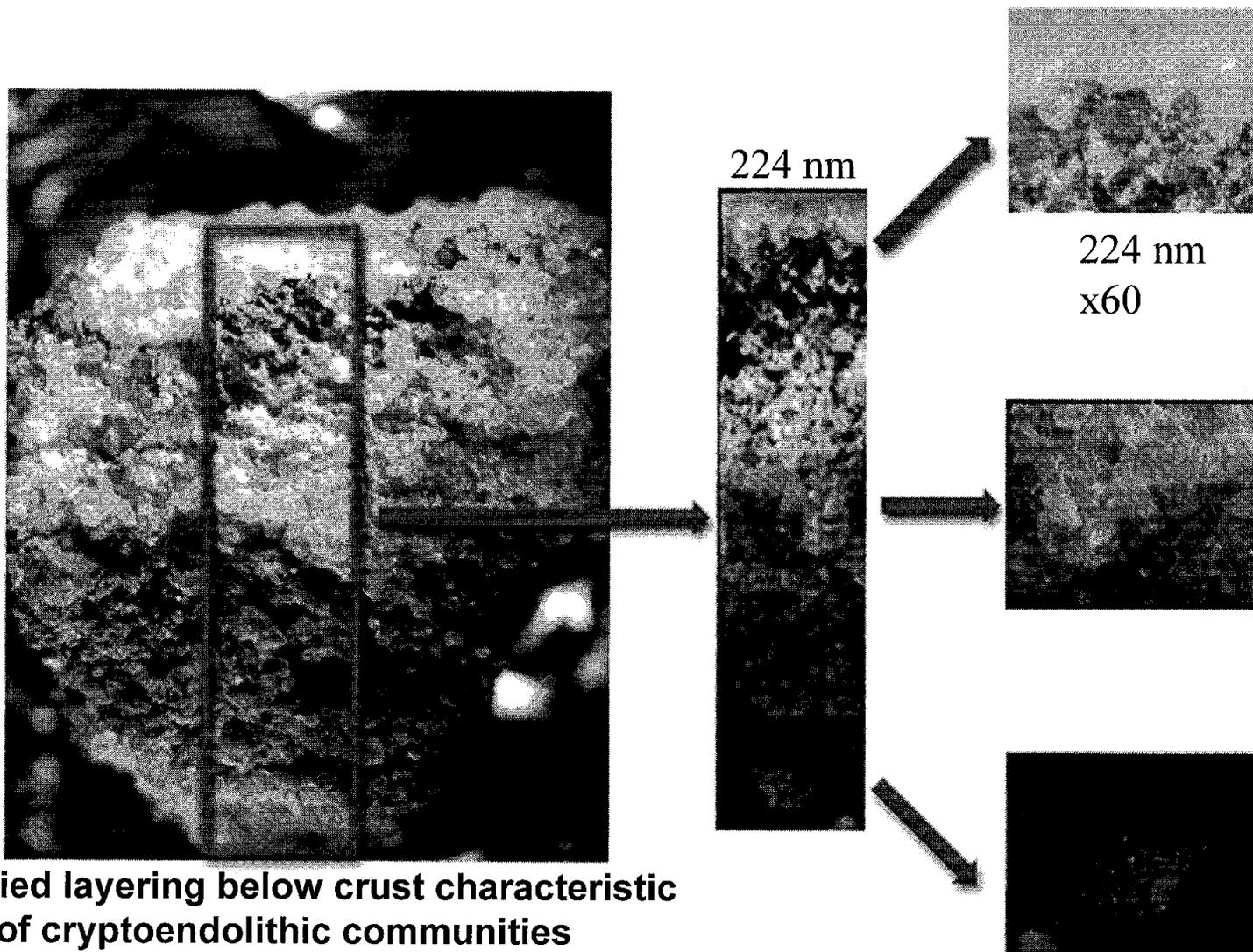
- **AFM can provide quantitative topographic imaging down to a few nanometers resolution in a modest resource envelope**
- **Chemical information is accessible through an IR-AFM combination well below a micron resolution**



Laser-induced fluorescence in microbial colonies



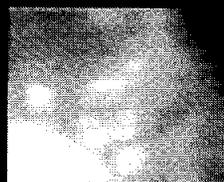
224 nm wavelength induces native fluorescence activity in aromatic amino acids.



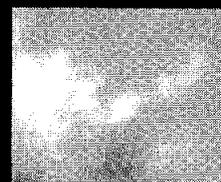
Deep Ultraviolet Laser Induced Fluorescence and Raman Spectroscopy of the Algal Community within an Antarctic Rock

Fluorescence
(Magnification: 300x)

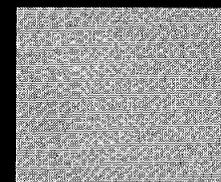
325nm



248nm

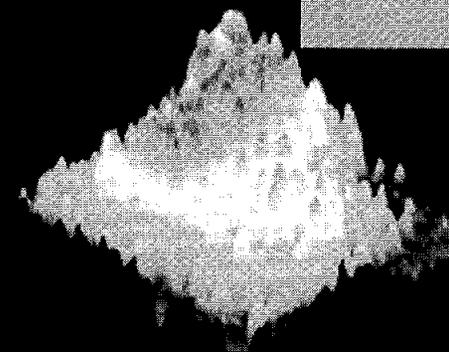
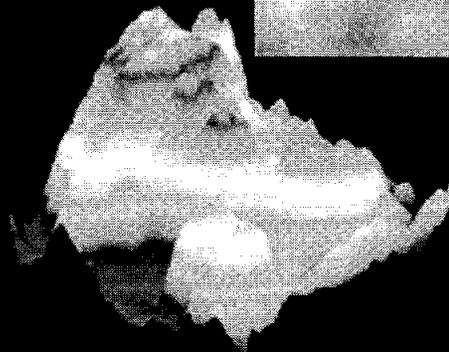
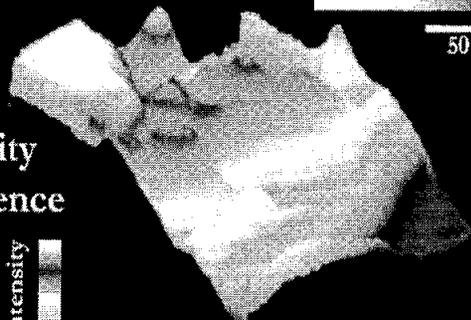


224nm

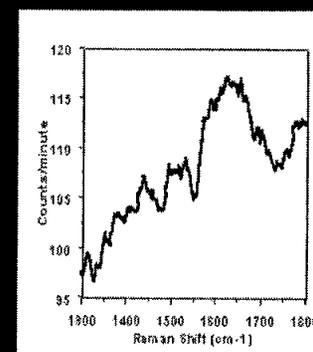
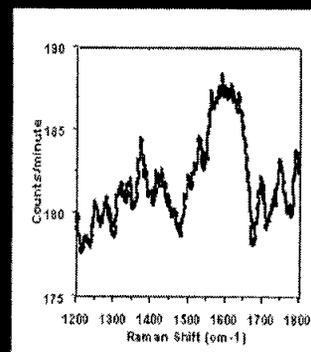
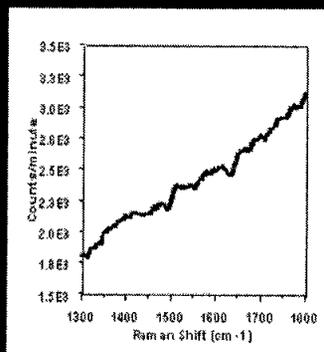


50 μ m

Pixel Intensity
of Fluorescence



Raman Spectra





Portable UV fluorescence imaging & Raman



20 cm X 25 cm X 50.8 cm

10 kg

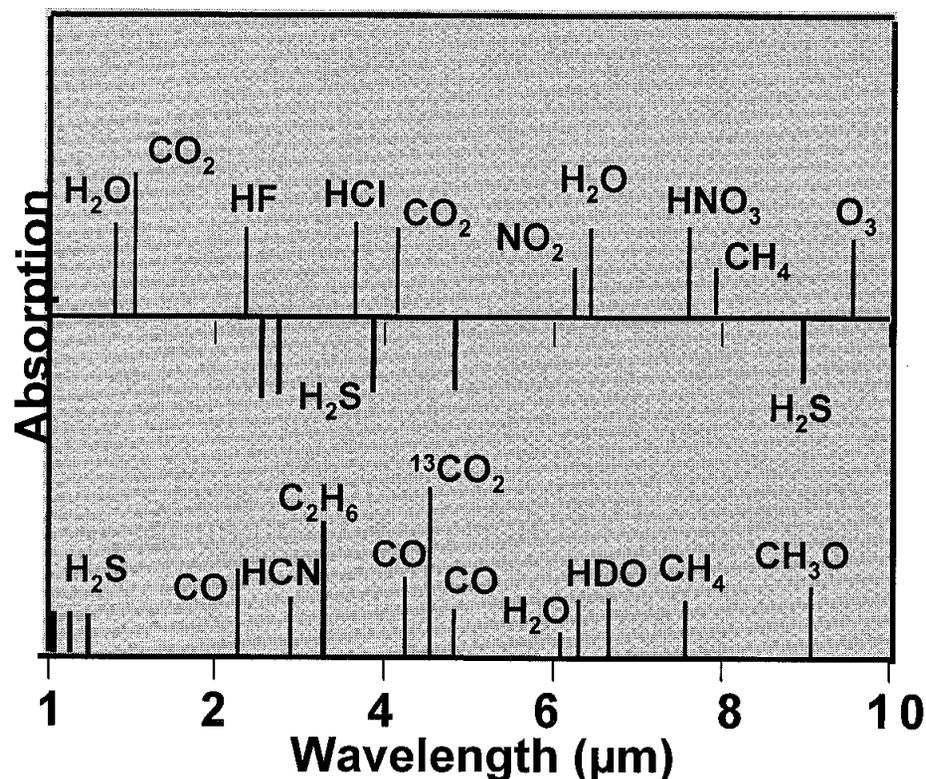
224 nm, 248 nm, 325 nm

< 100 W

P. Conrad/JPL
W. Hugs/Photon Systems



Laser-based instruments that cover 2–12 μm will enable the detection of all gases & volatiles of interest for planetary exploration, including isotopes—



- ◆ Gas evolution from rock, soil, or ice samples (e.g. CH_4 , NH_3 , HCN , HCL_3 , CO_2)
- ◆ Atmospheric composition
- ◆ Isotopic ratios $^{13}\text{C}/^{12}\text{C}$, $^{18}\text{O}/^{16}\text{O}$, $^{17}\text{O}/^{16}\text{O}$, $^{15}\text{N}/^{14}\text{N}$, $\text{H}_2\text{O}/\text{HDO}$
- ◆ ppb sensitivities

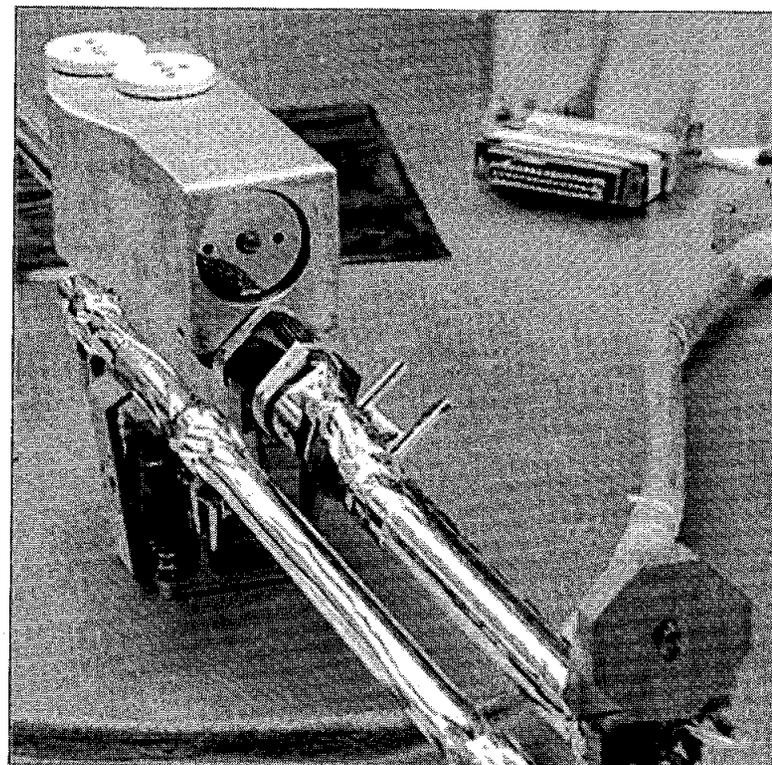
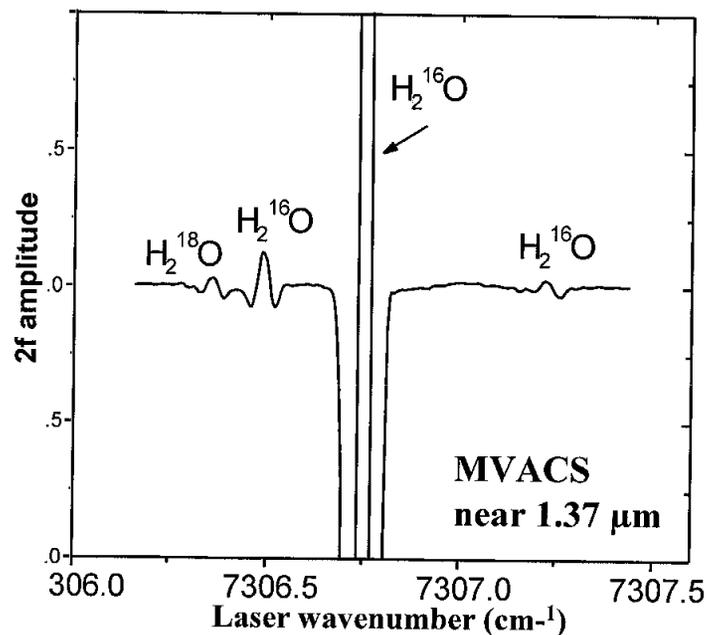


Laser spectroscopy first on MPL

JPL

MVACS carried 4 TDLs

- Meteorology package to measure water content of Mars atmosphere
1.37 μm (H_2O , 1 ppm)
- Thermally Evolved Gas Analyzer (TEGA) to measure volatile components of soil
1.37 μm & 2.04 μm (CO_2 isotopes)



Miniature laser spectrometer for the measurement of CO_2 & H_2O isotopes in the Martian atmosphere.

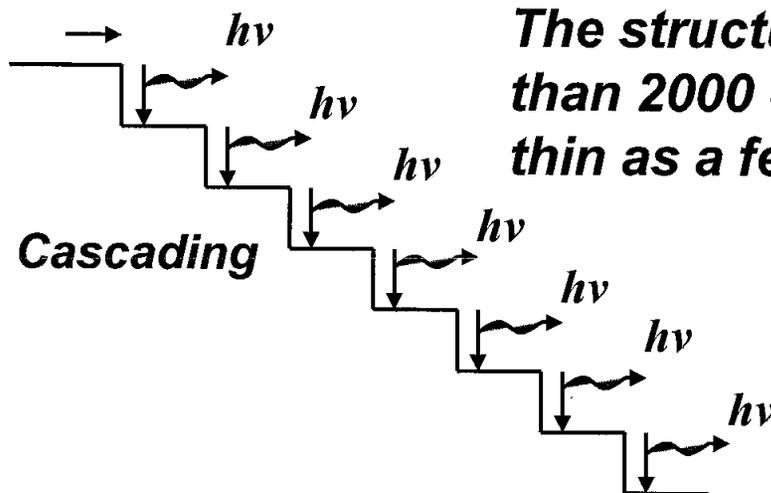
DS2 penetrator probe

- to measure subsurface water in Mars soil

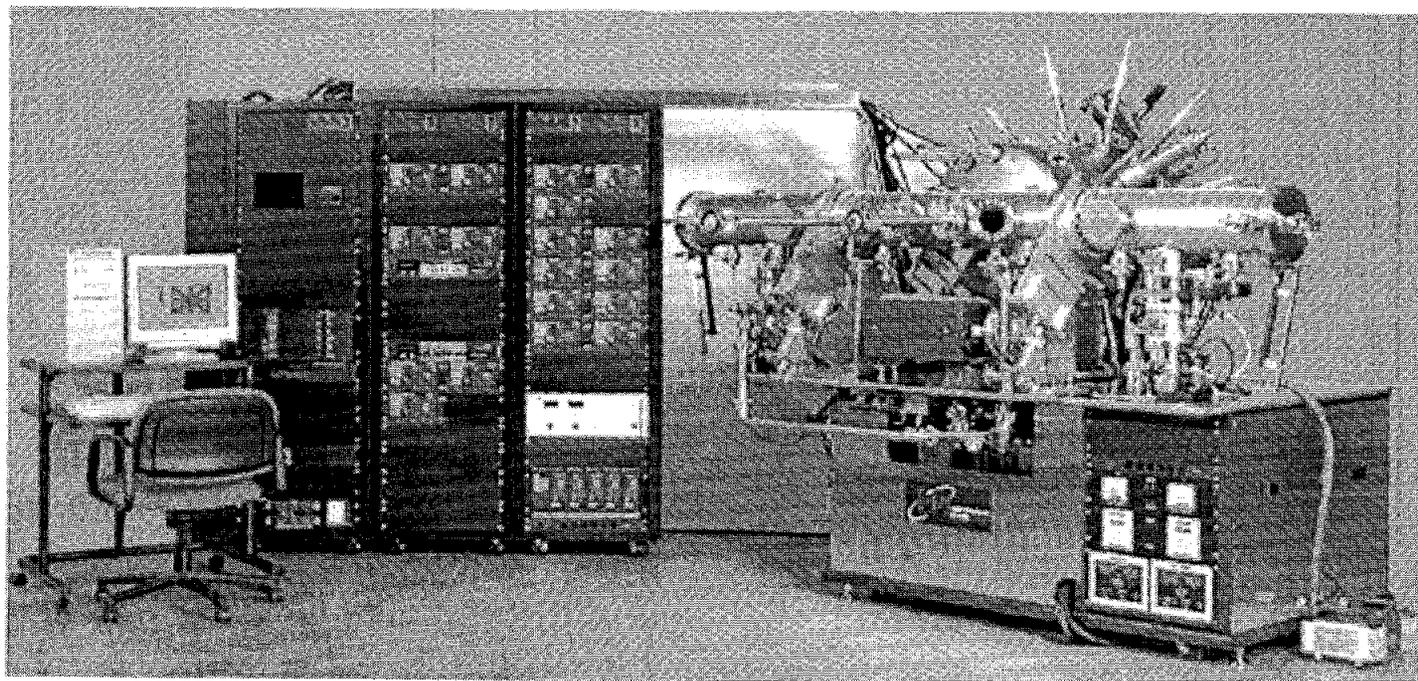


2–10 μm Interband cascade lasers

JPL



The structure is complicated—more than 2000 epitaxial layers, some as thin as a few atomic layers!





Sample Preparation



Examples:

EXPERIMENT	ANALYTICAL TOOL	SAMPLE HANDING/PREP
Viking biology	Scintillation, GCMS	Soil sieved, solution added, heated
MVACS (TEGA)	Molecular absorption spectroscopy	Soil sieved, heated
DS-2	Molecular absorption spectroscopy	Soil heated
MSL	Raman, Mossbauer (examples)	Rock grinder/sorter
'01 MECA	Electrochemistry (canceled)	Solution & calibrants added, stirring

- ◆ Our in situ instruments flown to date (or about to fly) have involved sensor systems that require no or simple, minimal sample handling & preparation
- ◆ Scooping or drilling used to deliver sample to instrument.



Future Sample Handling

JPL

EXAMPLE ANALYTICAL TOOLS

Emerging “on-chip” detectors

Biosensors

Electrochemical

Integrated electro optical

Polymeric artificial antibodies

Emerging micro analytical tools

SAMPLE HANDING & PREPARATION

Wet chemistry procedures

Separations

Reactions, derivatizations

Reagent preparation

Volumetric metering

Mixing

Fluid positioning

Filtering, concentrating,

Heating, cooling

Washing, storage...

Dry preparation procedures

Cutting, polishing, sectioning

*Separation by physical & chemical
properties*

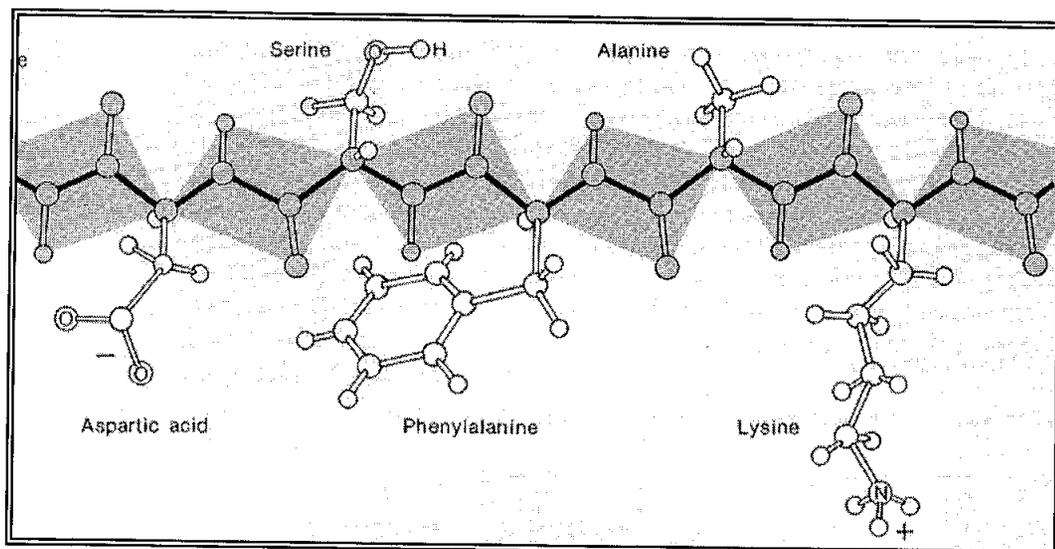
Sublimation

Need increasingly sophisticated miniaturized lab-bench procedures for “dry” and “wet” micro analytical systems



Looking for Amino Acids

JPL



- ◆ They are readily synthesized under plausible prebiotic conditions
- ◆ Long lifetime under dry, cold conditions
- ◆ They are abundant in carbonaceous chondrites
- ◆ Molecular architecture can be used to distinguish abiotic versus biotic origins

◆ Abiotic chemistry

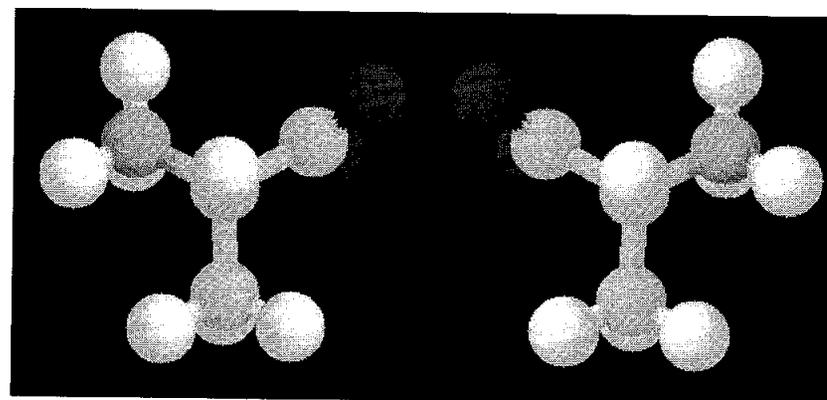
- Presence of non-protein amino acids and racemic amino acids

◆ Extinct life

- Amino acids with different amounts of racemization

◆ Extant life

- Amino acid homochirality



L-alanine

D-alanine



Composition and Chirality Analysis through CE

JPL

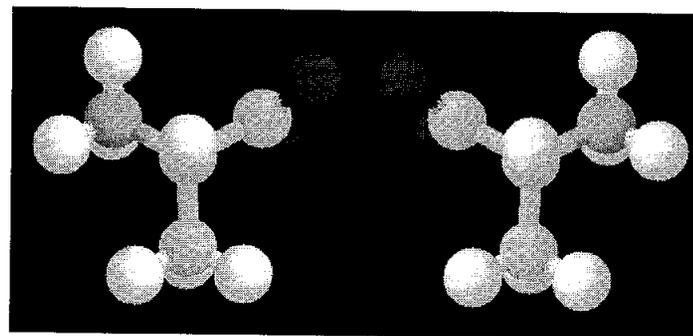
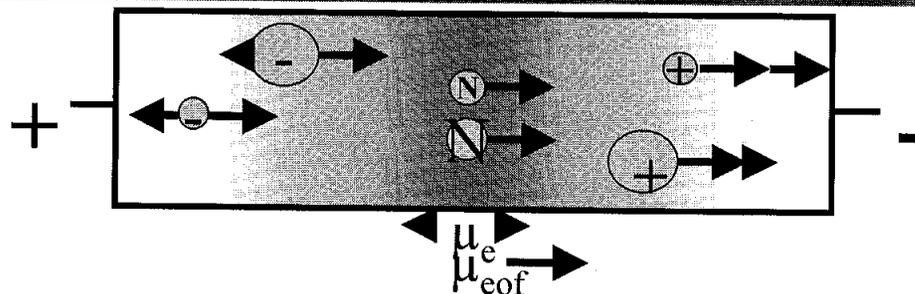
- In CZE, EOF sweeps all molecules to the cathode
- CE separates amino acids based on charge/size ratio
- L and D-amino acids are not resolved because of identical mobilities
- Cyclodextrins provide enantiomeric resolution of amino acids because of different complexation constants and altered size when complexed:



$$K_L \neq K_D$$

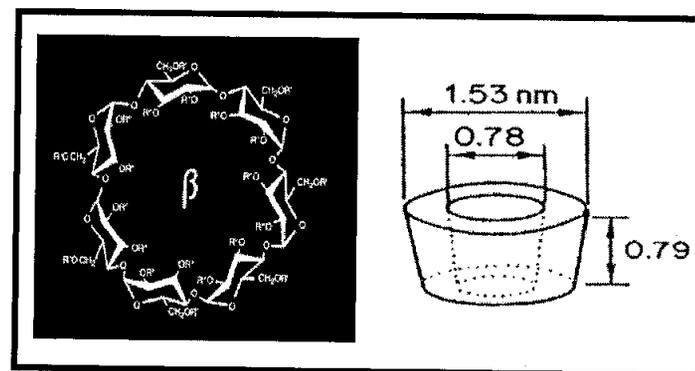
- The net apparent mobilities μ_a are different

$$\mu_L \neq \mu_D$$



L-alanine

D-alanine



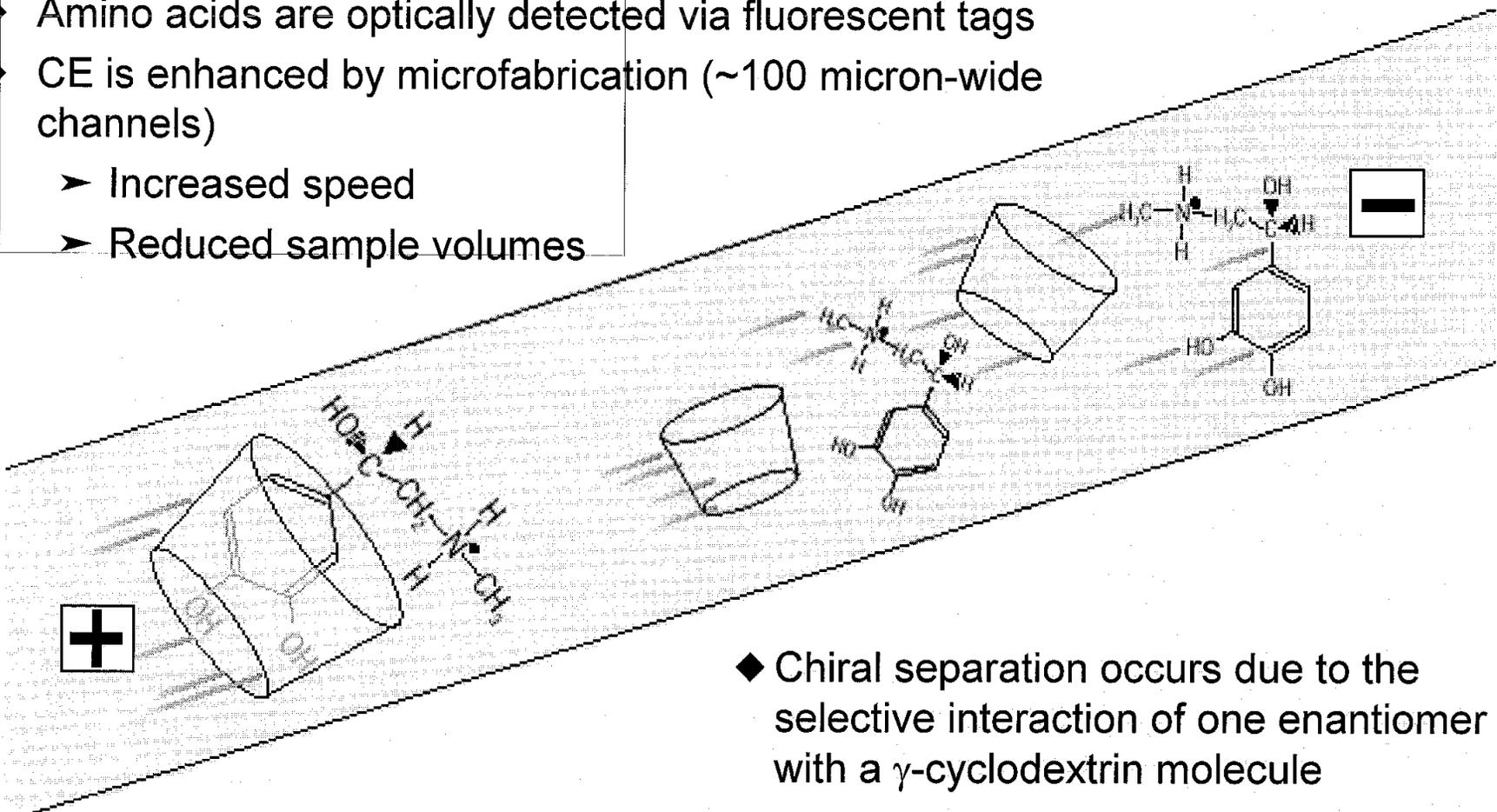
cyclodextrin



μ Capillary electrophoresis

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- ◆ Amino acid composition and especially chirality is a possible probe for extinct or extant life in extraterrestrial environments.
- ◆ Microfabricated devices are now available that can determine amino acid composition and chirality in minutes on only nanoliters of sample.
- ◆ Amino acids are optically detected via fluorescent tags
- ◆ CE is enhanced by microfabrication (~100 micron-wide channels)
 - Increased speed
 - Reduced sample volumes

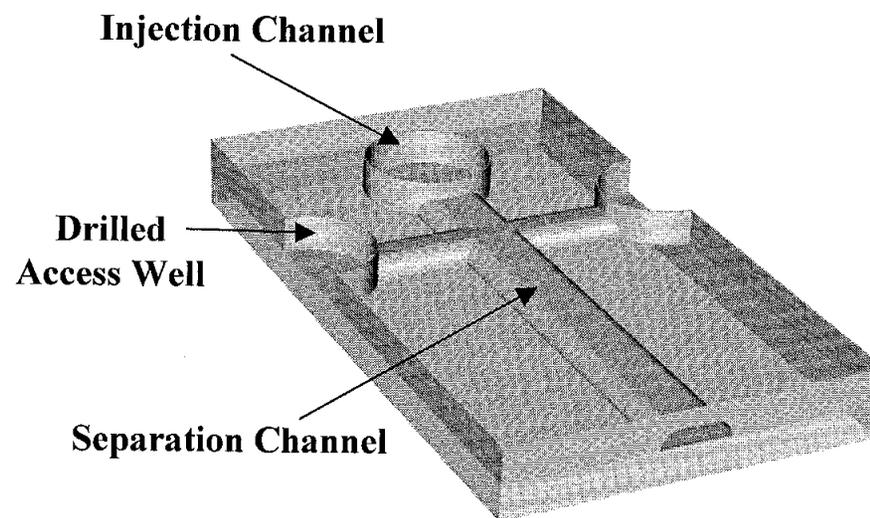
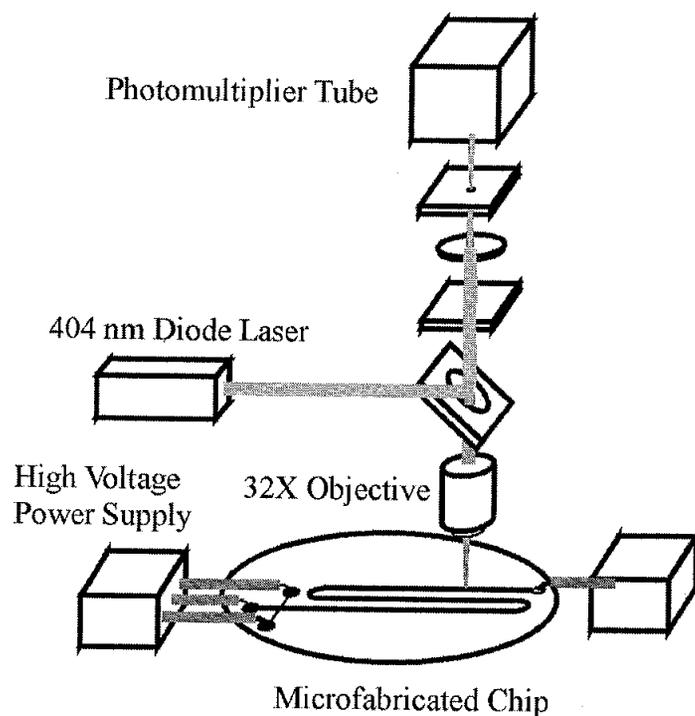


- ◆ Chiral separation occurs due to the selective interaction of one enantiomer with a γ -cyclodextrin molecule



Separation and Detection System

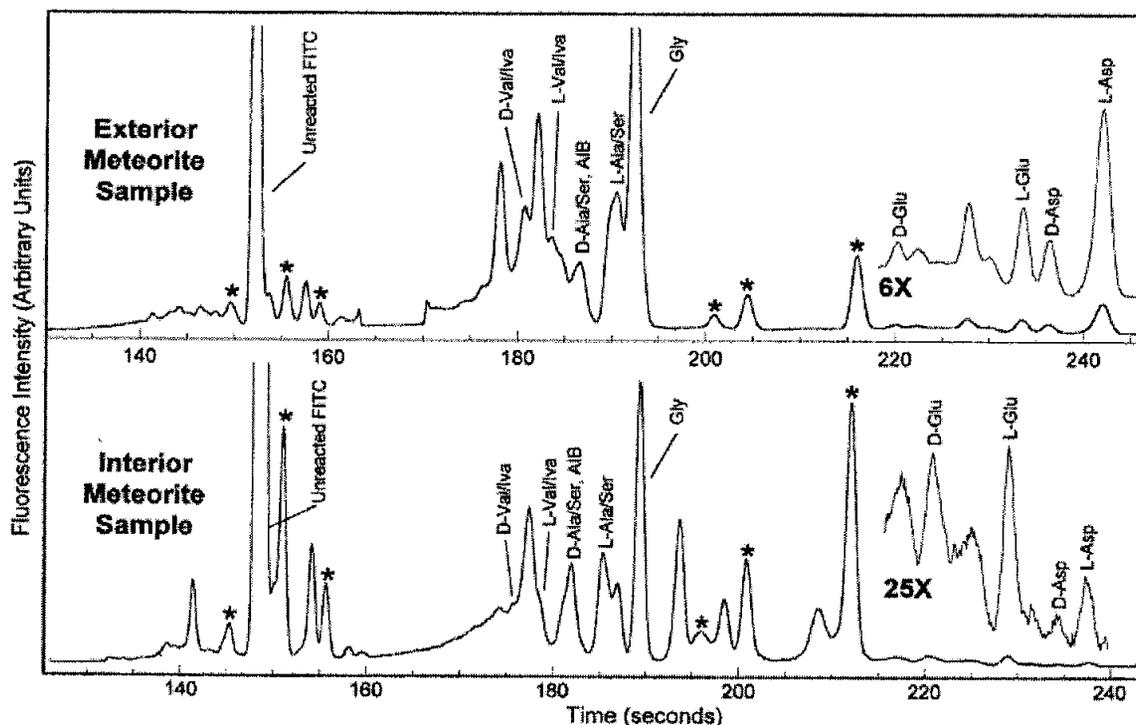
JPL



- ◆ The test system consists of a folded electrophoresis channel (19.0 cm long x 150 mm wide x 20 mm deep) photolithographically fabricated in a 10 cm-diameter glass wafer sandwich.
- ◆ A laser-excited confocal fluorescence detection apparatus provides sub-attomole ($<10^{-18}$ mole) sensitivity {Hutt et al. *Anal. Chem* 71, 4000 (1999)}.



μ CE analysis of Murchison Meteorite



Murchison Asp and Glu D/L Ratios

Asp = aspartic acid

Glu = glutamic acid

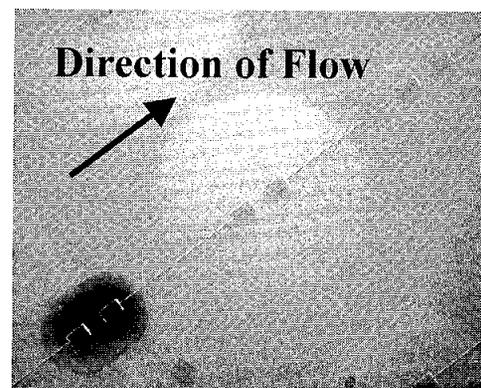
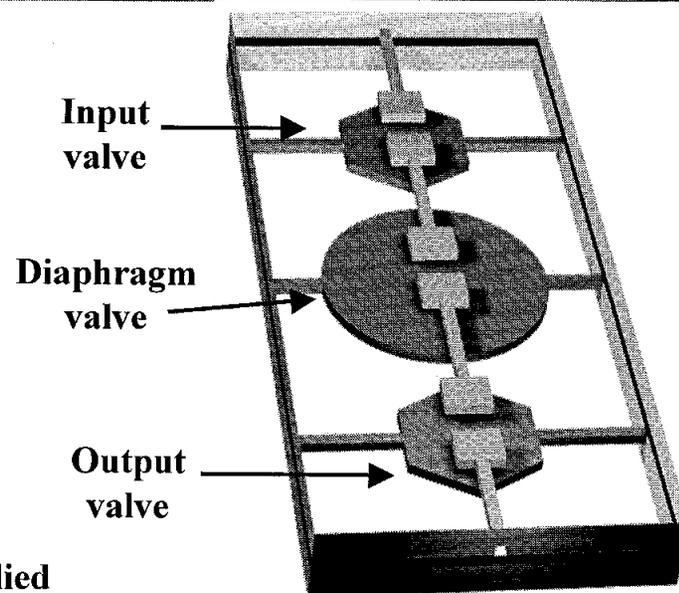
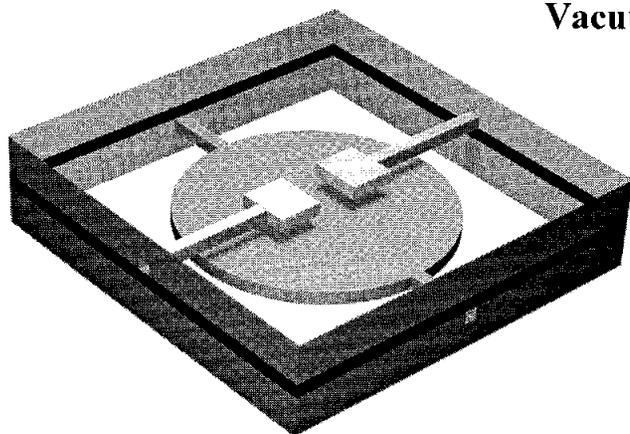
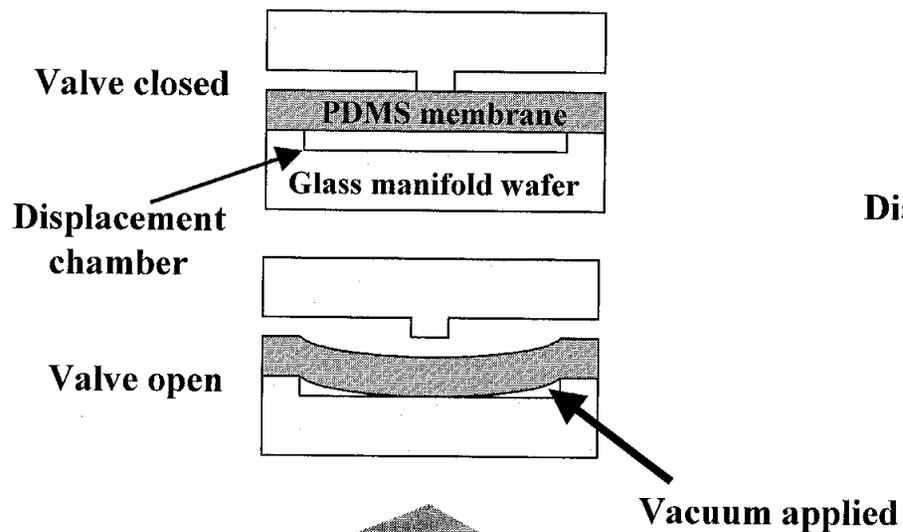
<u>Amino acid</u>	<u>microchip</u>	<u>HPLC</u>
Glu exterior	0.33 ± 0.04	0.3 ± 0.1
Glu interior	0.65 ± 0.07	0.7 ± 0.1
Asp exterior	0.21 ± 0.03	0.3 ± 0.1
Asp interior	0.30 ± 0.06	0.3 ± 0.1



Microfabricated Valves and Pumps



Side View



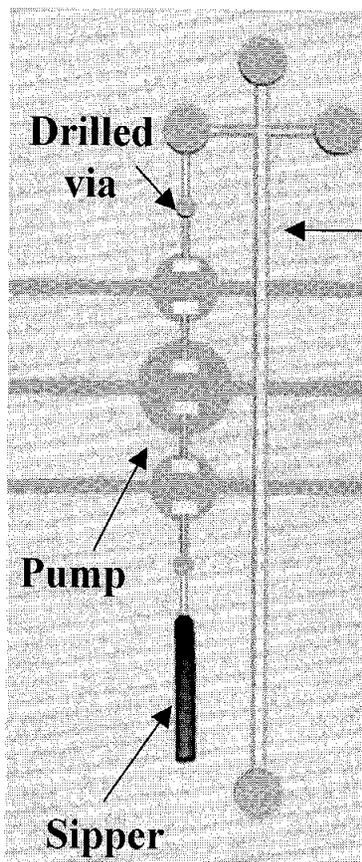
- Pump 10 nL to 10 uL per actuation
- Flow rates from 1 to 150 nL/s
- Self-priming, pump against head pressures



Sample collection to separation



Top



Side

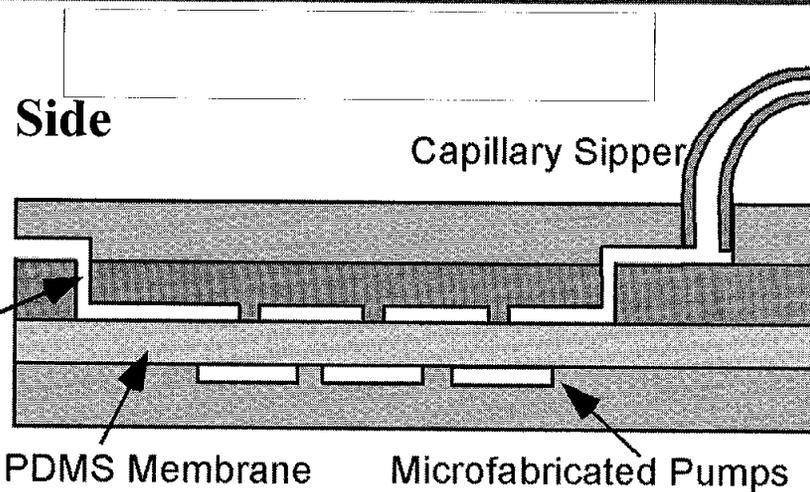
To separation channel

Via

Capillary Sipper

PDMS Membrane

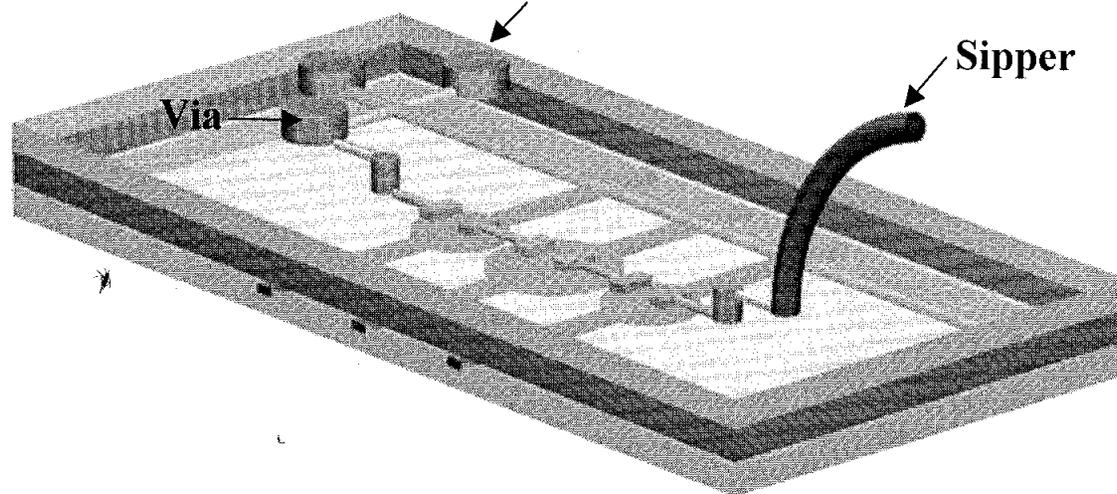
Microfabricated Pumps



Drilled access wells

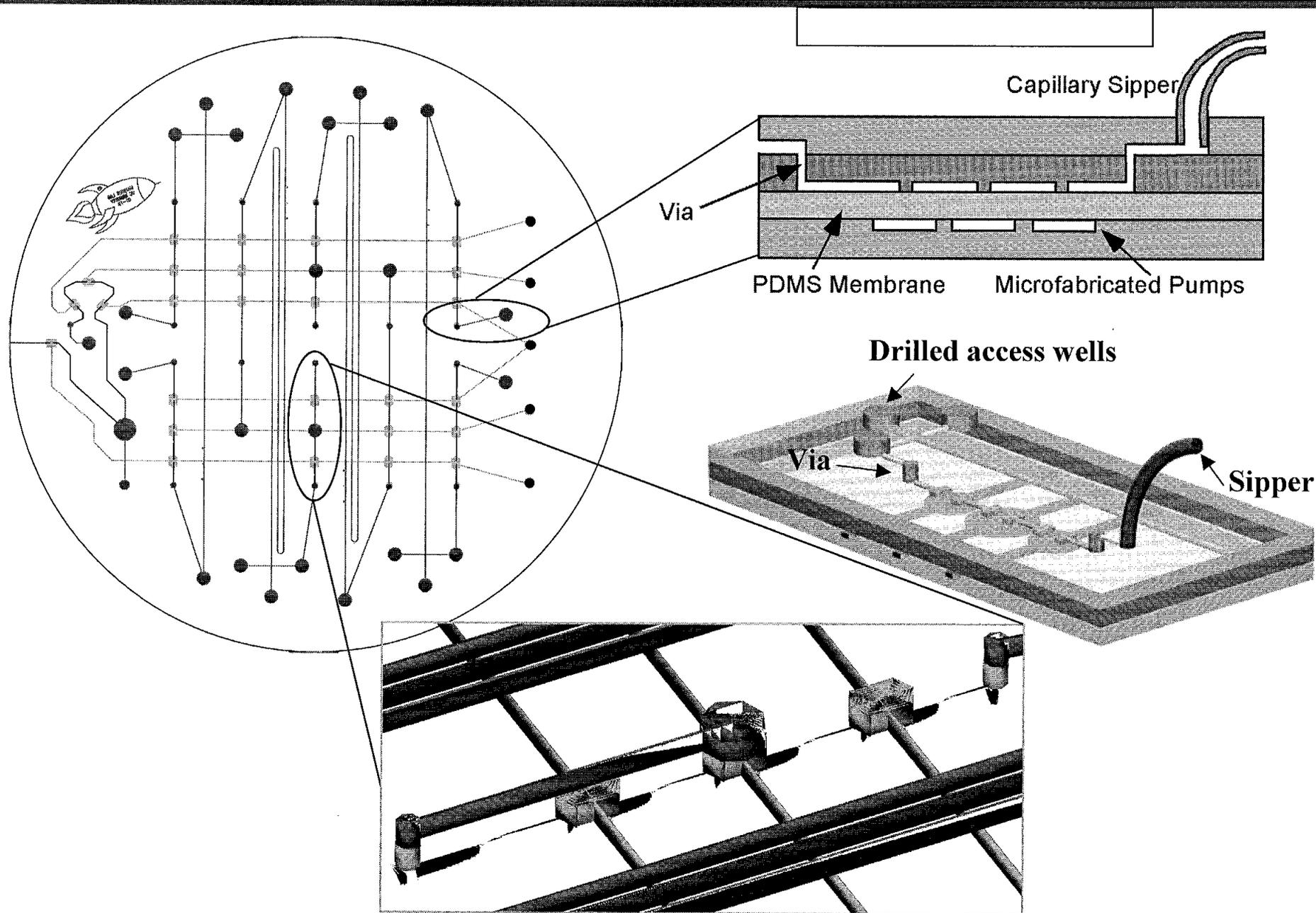
Via

Sipper





Sample collection to separation

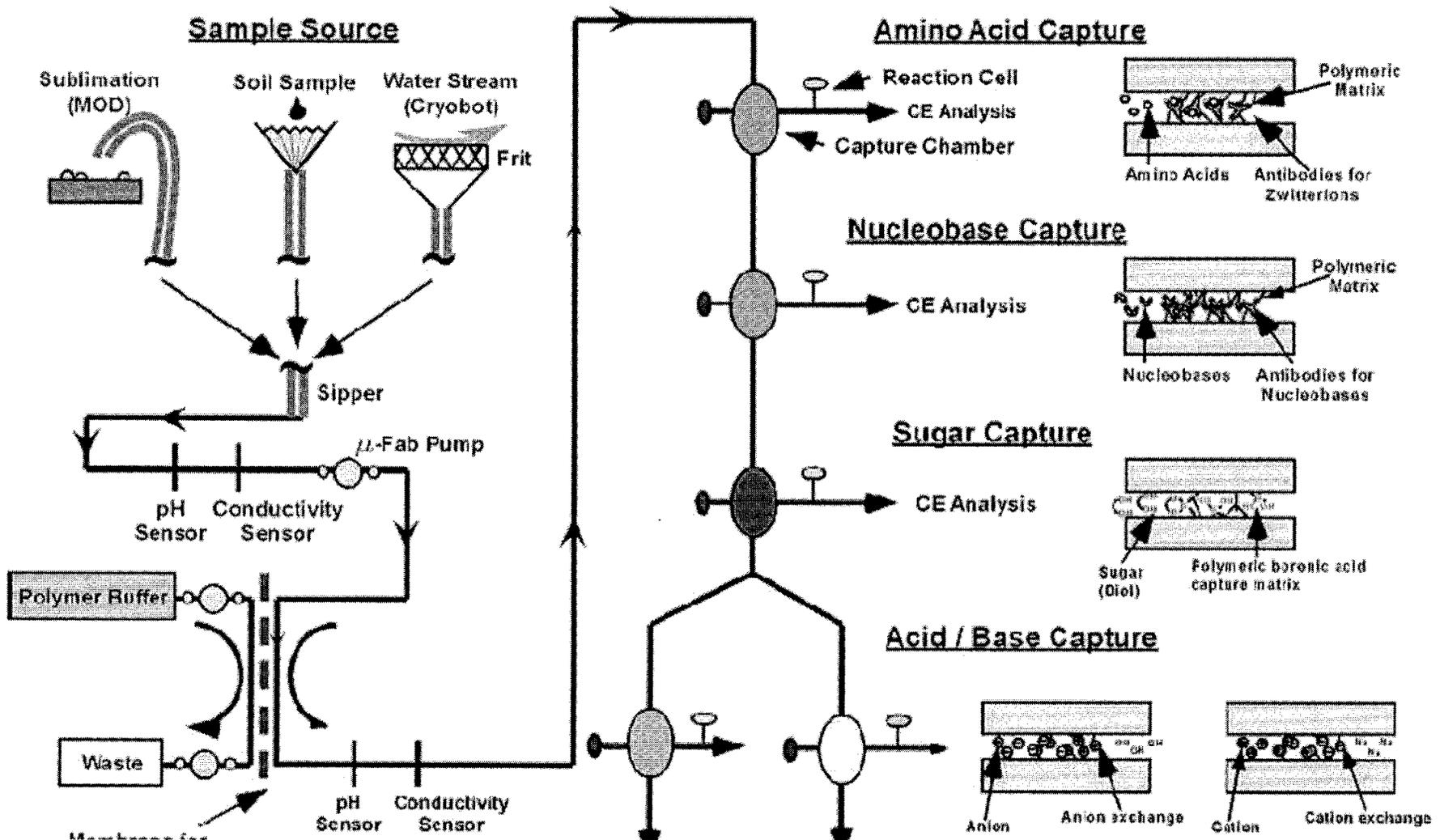




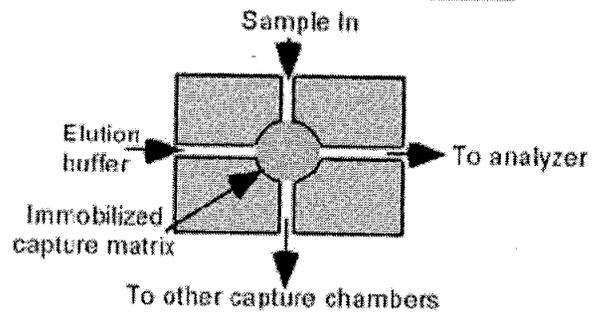
Lab-on-a-chip future concept

JPL

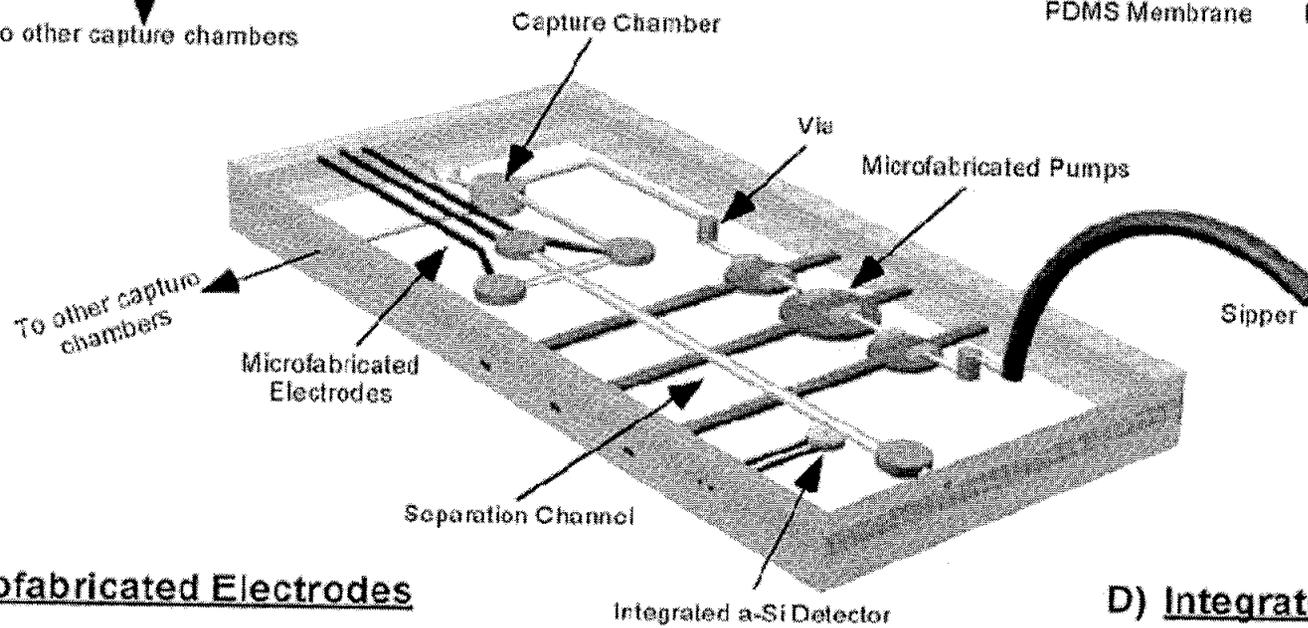
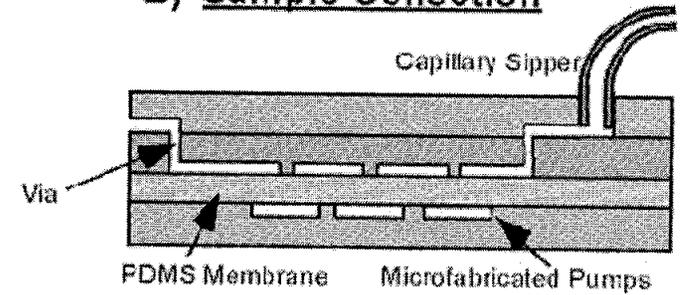
UCB/UCSD/JPL



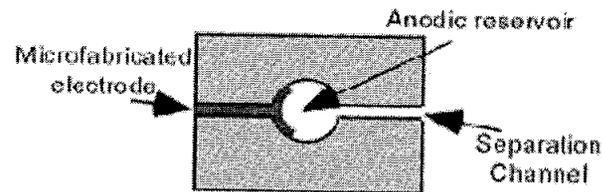
A) Capture Matrix Chambers



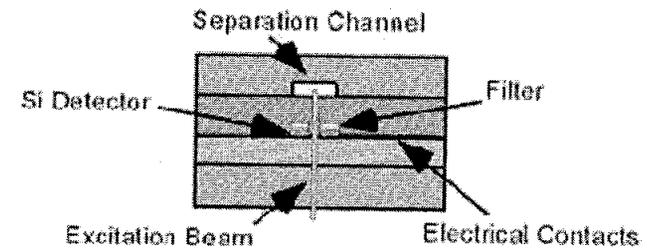
B) Sample Collection



C) Microfabricated Electrodes

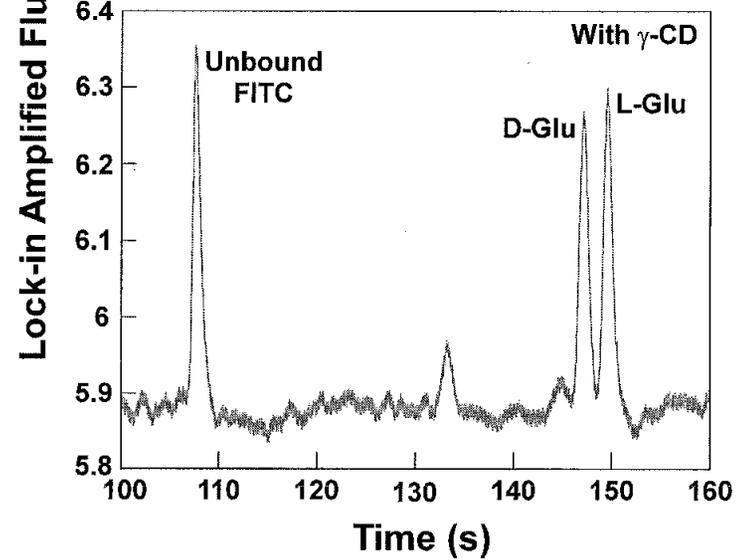
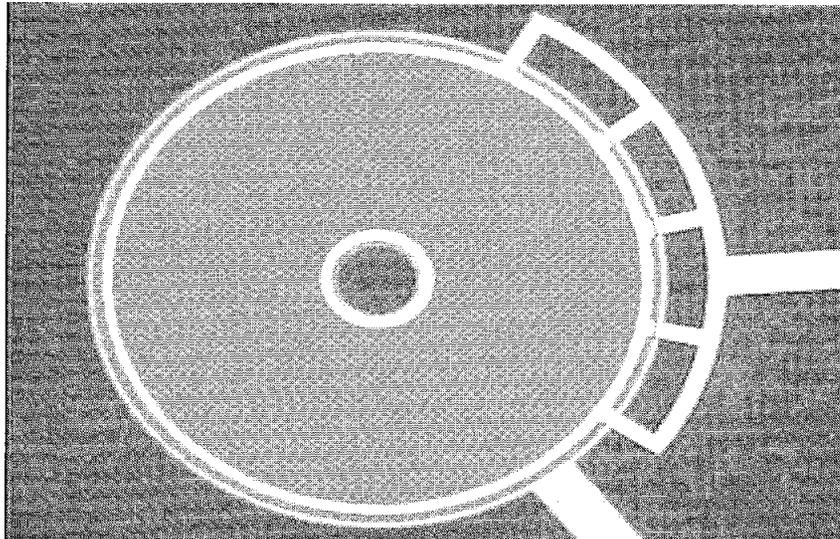
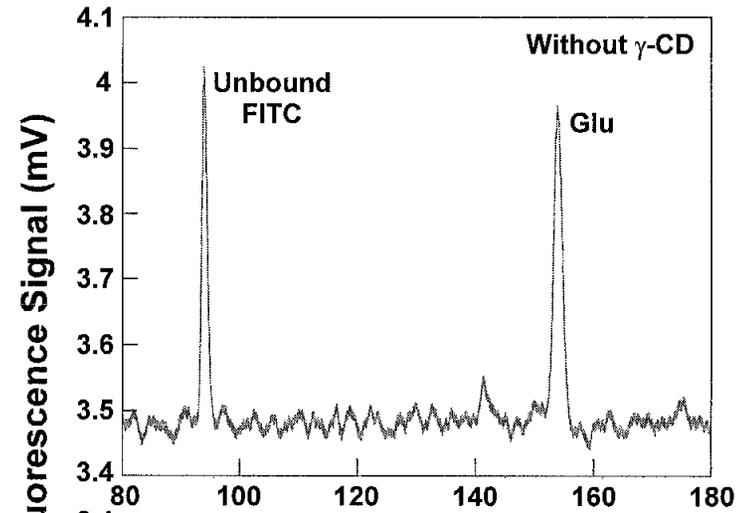
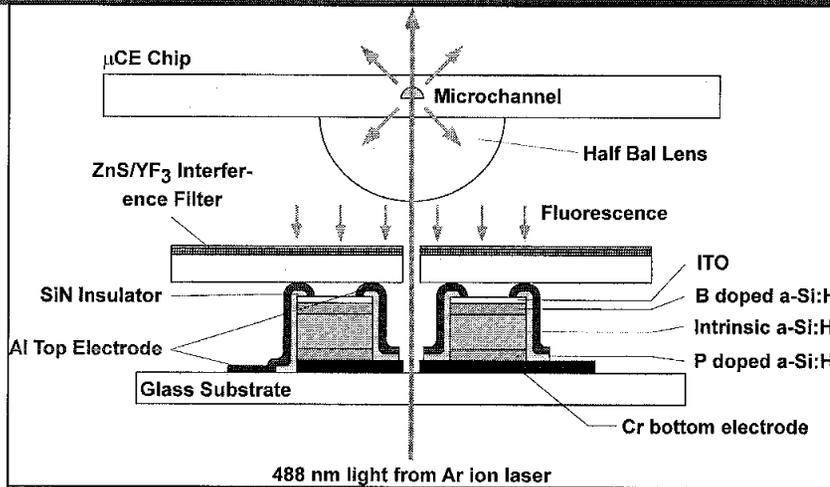


D) Integrated Detector





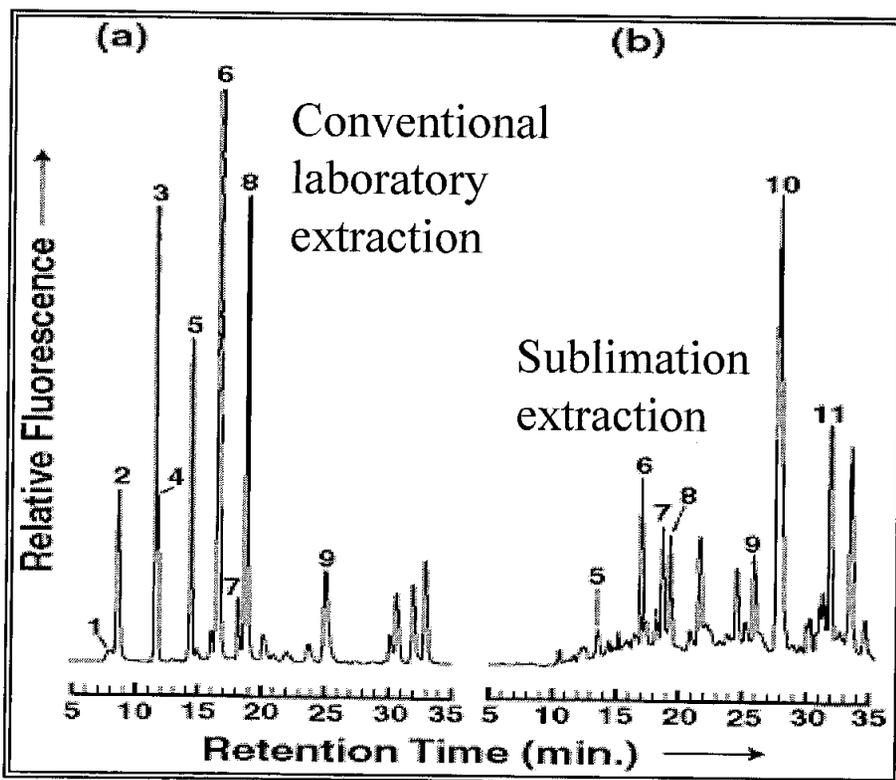
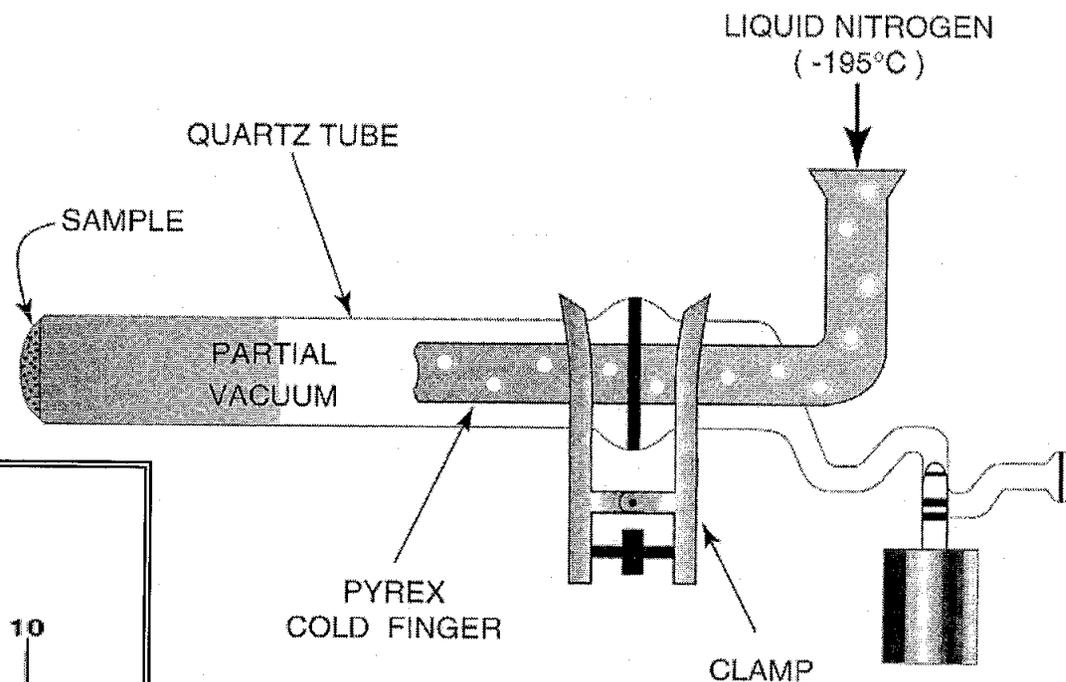
Amino Acid Analysis with Integrated Detector



Work performed by Dr. Toshihiro Kamei



MOD organic extraction

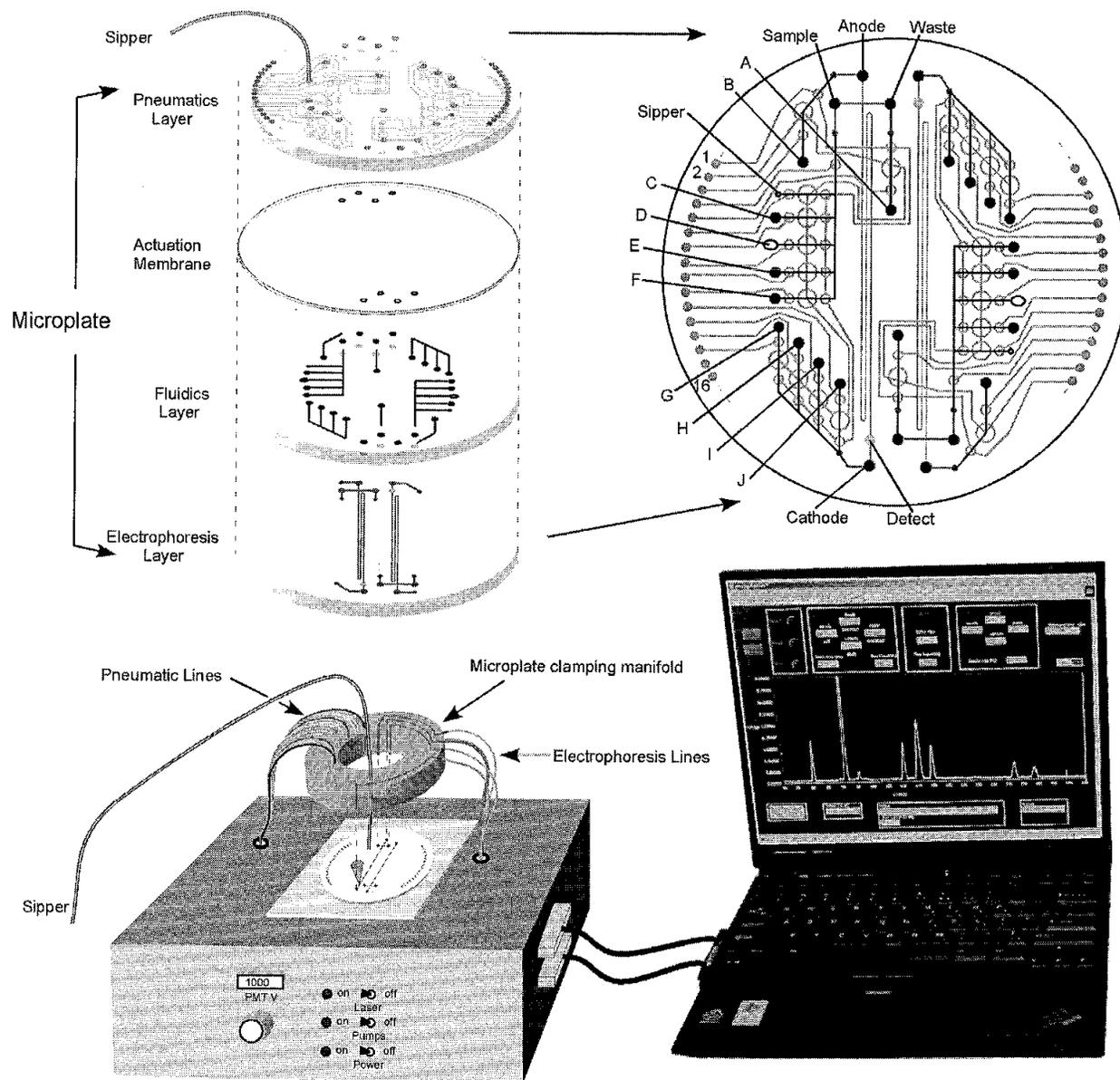


- ◆ Mars analog soil sample inoculated with *E. coli* cells
- ◆ 500°C for 30 seconds
- ◆ UCSD (J. Bada, PI)



Portable Amino Acid Analysis Devices

JPL



- ❖ 16 pneumatic lines for fluidic control
- ❖ Thermoelectric cooling
- ❖ Designed for either 404 nm (Fluorescamine) or 488 nm (FITC) diode laser
- ❖ Complete automated analysis for obtaining sample, labeling, diluting, and filling/flushing, CE channels for analysis

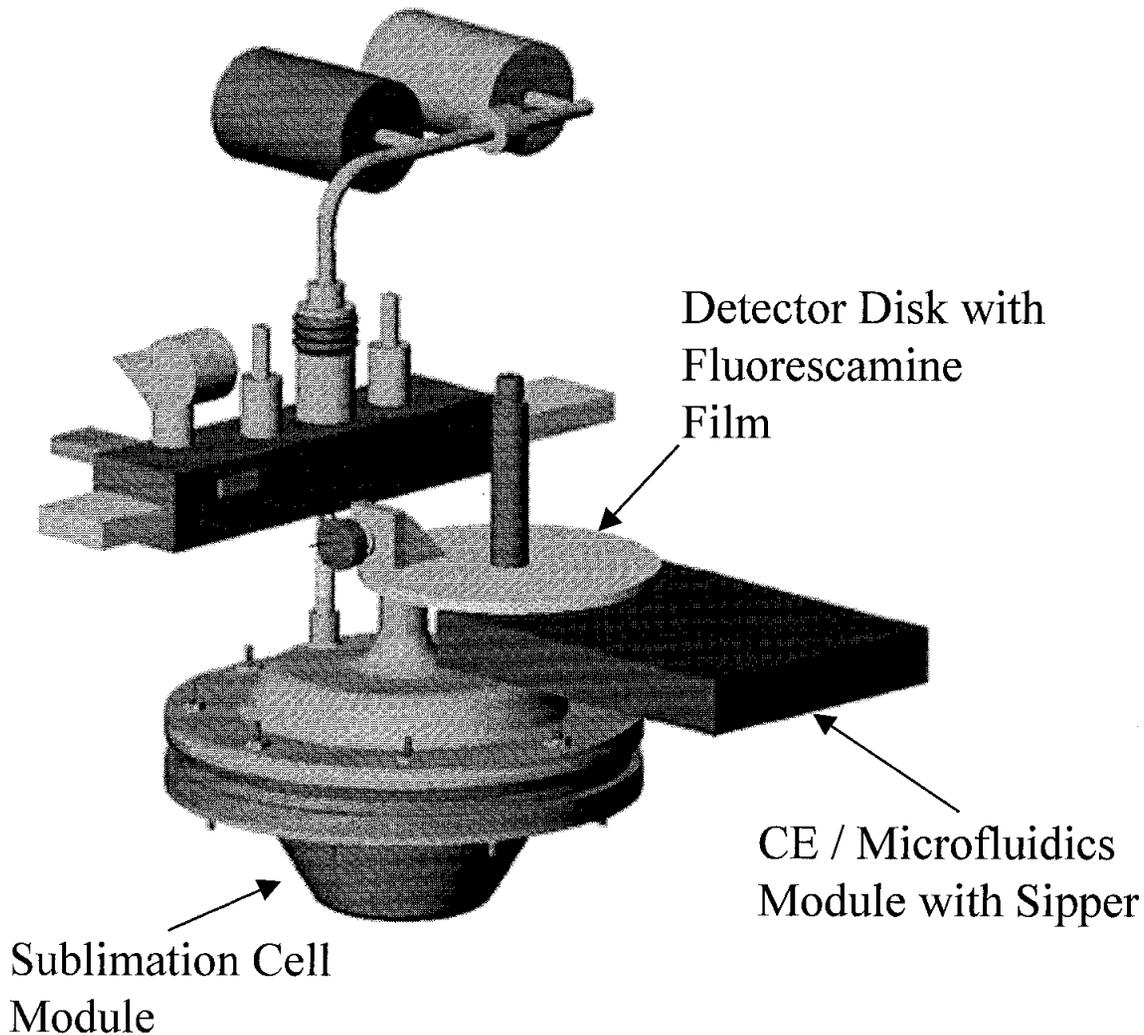
R. Mathies, UC Berkeley



Organic Analyzer for Life Detection

JPL

Systems are being developed that include highly complex sample preparation—



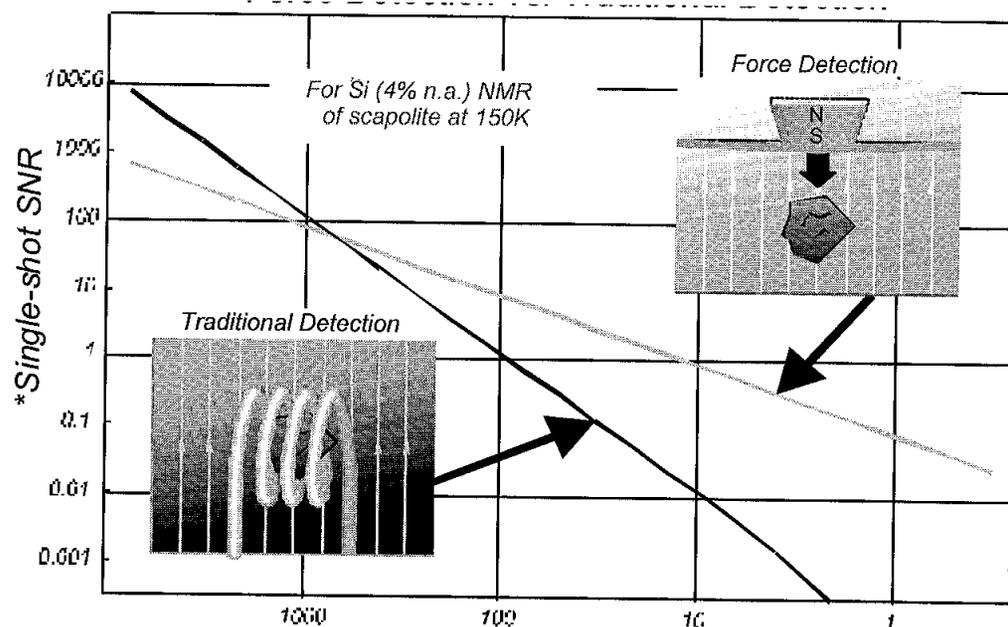
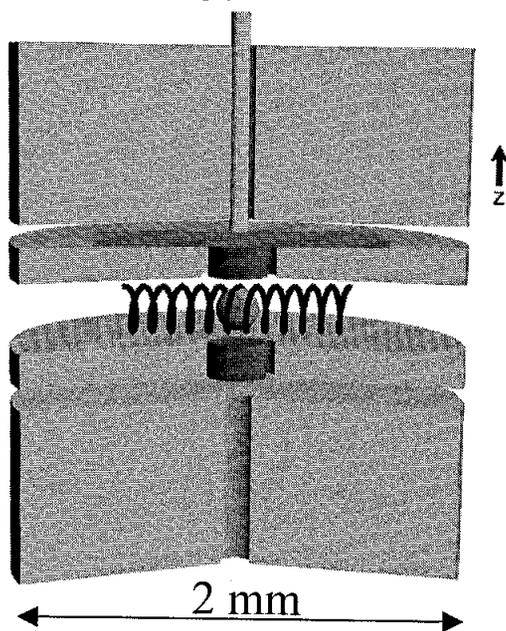
- ◆ Extracts, concentrates, fluorescently labels, and analyzes organic molecules
- ◆ Amino acids, amines, carboxylic acids, sugars, & nucleobases, including chirality
- ◆ 6 orders of magnitude more sensitive than Viking GCMS
- ◆ Martian soils, rocks, polar ice



Force-Detection NMR

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- ◆ 1 cm³ volume, < 100 mW RF, several ppt accuracy for all isotopes with nuclear magnetic moments
- ◆ RF-induced change in magnetic moment sensed by movement of small central magnet suspended on beam (milli angstrom movement sensed by optical interferometry)



- ◆ Resolve chemically distinct sites and bonding environments from NMR line positions, amplitudes, relaxation times, & coupling constants
- ◆ Hydrocarbons in water, solid samples & mineral phases (especially good with light elements such as H, Li, B, C, N, F, Cl, and S), H₂S and hydrates, microbiological sensing (proteins, functional groups)

D. Weitkamp (CIT), T. George (JPL)

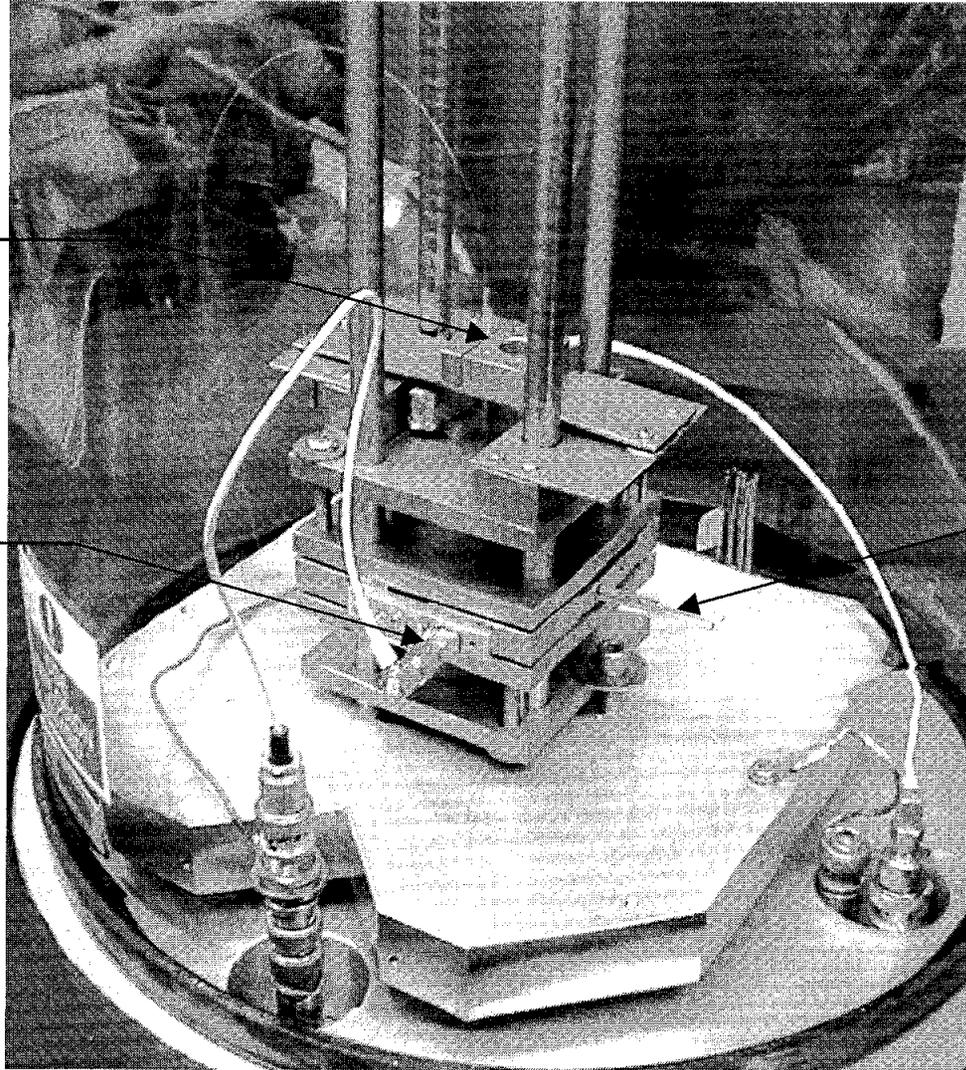


"Big Mac" laboratory test setup

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Fiber Optic

RF Coil



Proof-of-concept test setup for spectrometer which is 25x final size, tested in vacuum in a bell jar

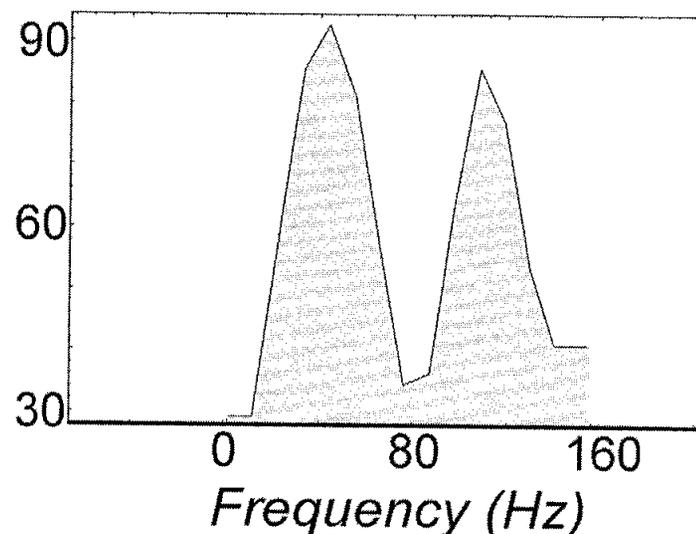
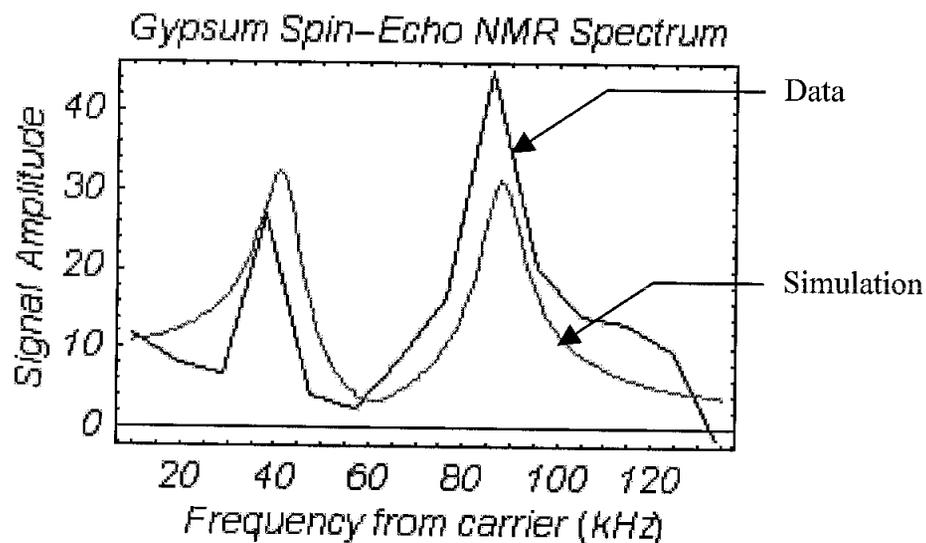
Sample holder

Prof. D. Weitekamp (Caltech)
Thomas George (JPL)



FDNMR proof-of-concept spectra

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- ◆ Using the “Bigmac” spectrometer, we have determined chemically specific material parameters that were previously not achievable via force detection techniques.
- ◆ Spectrum at left shows a splitting due to the intramolecular dipole-dipole coupling of hydrogens in the waters of hydration of a gypsum crystal.
- ◆ Spectrum at right shows a splitting, at much higher resolution, due to the ^{19}F - ^1H through-bond, or “J”, coupling in liquid fluoroacetonitrile (CF_2HCN).

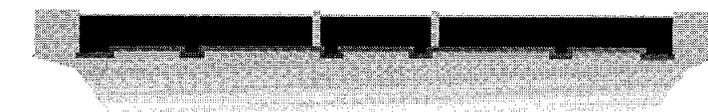
Prof. D. Weitekamp (Caltech)
Thomas George (JPL)



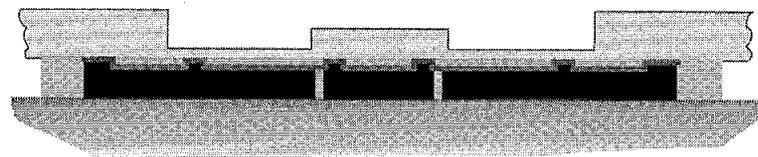
1-step plating fabrication process



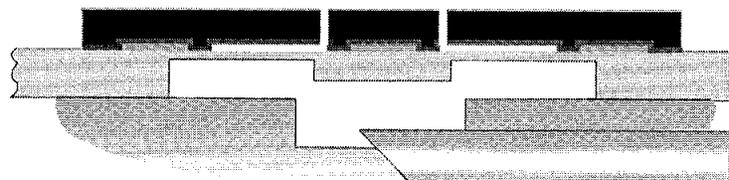
1. Thermal oxidation and patterning for sacrificial layer.
2. Deposit Cr/Au (200Å/1000Å) plating seed layer and pattern photoresist mold.
3. Electroplate ring and detector magnets 10 μm thick.
4. Protect front side by wax-mounting to wafer.
5. Pattern back and create stress buttress and oscillator beam using deep RIE.
6. Remove sacrificial oxide (BOE).
7. Bond field magnet and fiber to back.



Steps 1-3

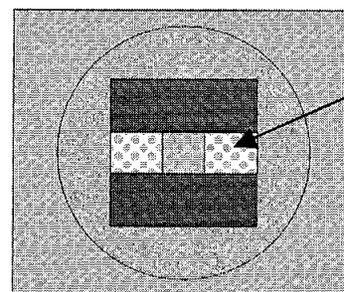


Steps 4,5



Steps 6,7

- | | |
|---|---|
|  silicon |  protect wafer |
|  plated magnet |  field magnet |
|  oxide |  photoresist |
|  seed layer |  fiber |



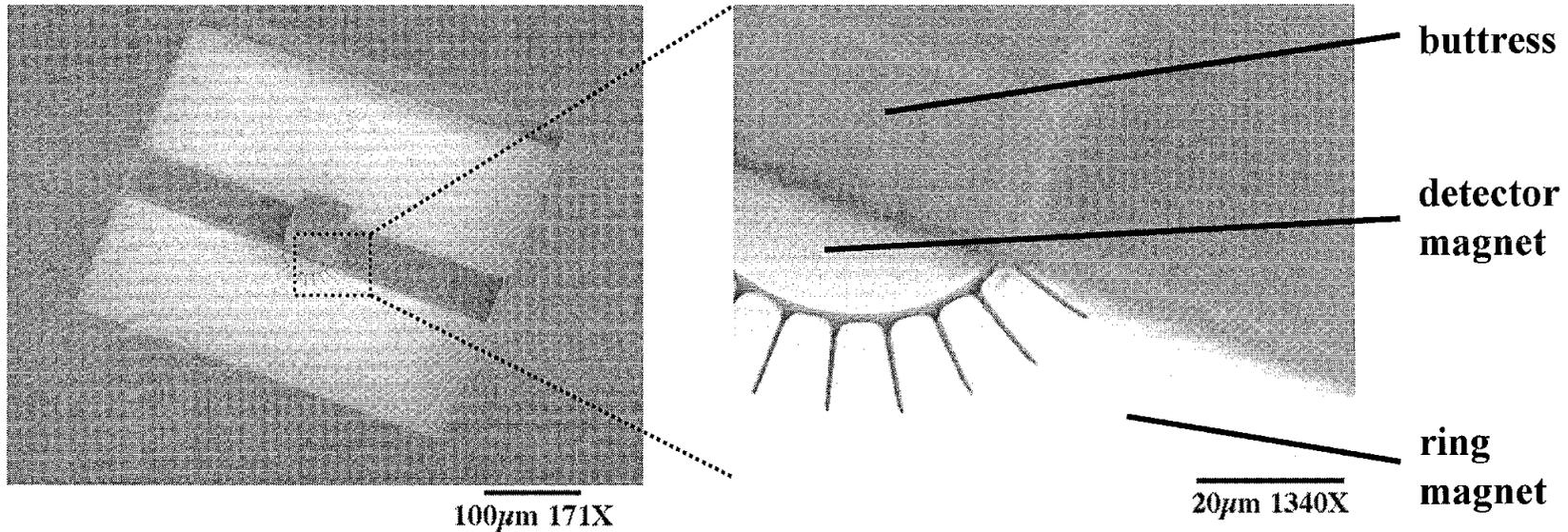
Oscillator Beam

Backside Plan View

Prof. D. Weitekamp (Caltech)
Thomas George (JPL)



Deep-RIE-defined beam and buttress with plated magnets



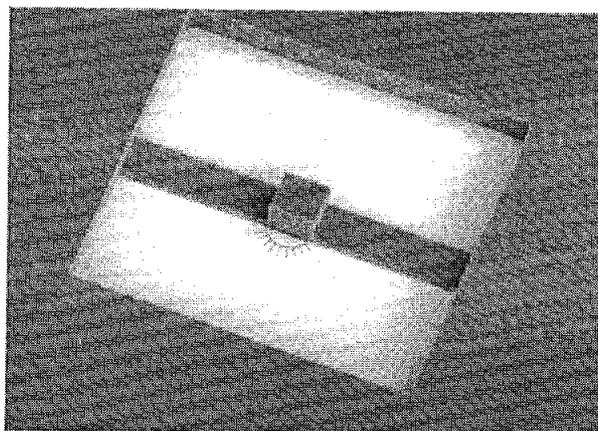
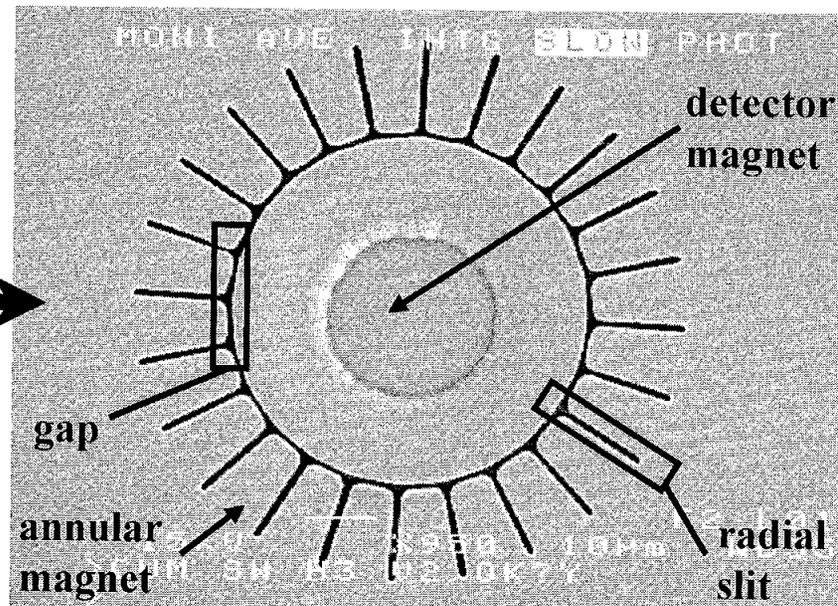
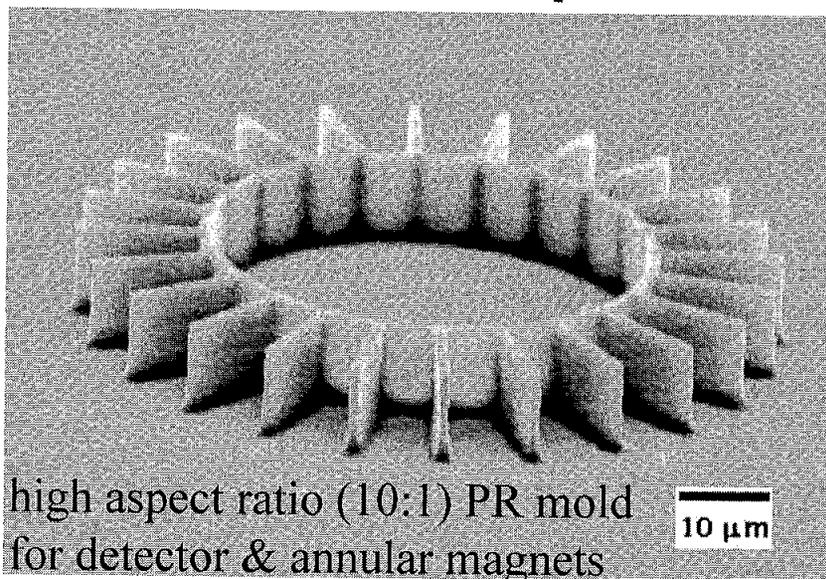
- ◆ Deep RIE process used to define 5 μ m-thick Si beam and stress buttress on backside of plated magnet array
- ◆ Detector/ring magnet gap is \sim 1 μ m with slits for eddy-current reduction
- ◆ After release of PR and oxide sacrificial layers, magnets are still intact and not stress curled.



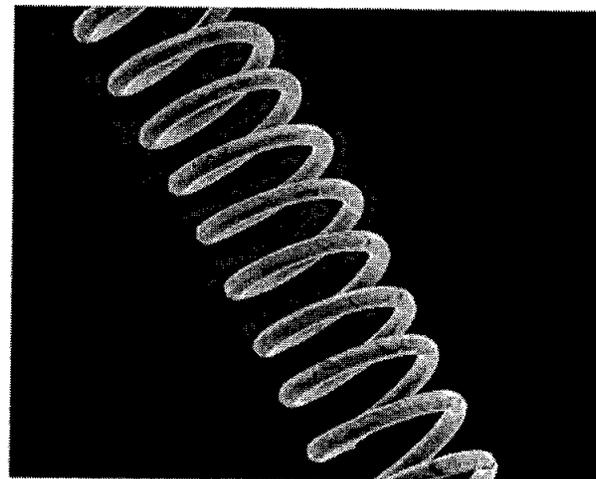
MEMS FD-NMR



Nearing completion of microfabricated detectors for MEMS FD-NMR spectrometer—



100 μm 171X



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Questions?