



National Aeronautics and Space Administration

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Pasadena, California

A Sensitive Real-Time Spore Detection Assay

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Abstract

NASA monitors the cleanliness of spacecraft to limit terrestrial microbes being transferred to other planetary bodies. Spore-forming organisms are of particular concern. The current NASA standard procedure involves collecting spacecraft surface sample swabs and detecting spores using a culture-based assay. This traditional method is laborious and requires a long incubation time. A sensitive, rapid, and real-time spore detection (RSD) assay is needed to meet demanding spacecraft assembly schedules. A highly sensitive ATP bioluminescence assay has been developed to determine the microbial cleanliness of spacecraft and associated environments. This ATP assay is rapid and simple. However, the level of ATP is very low in spores and the ATP assay is not very sensitive for spore detection. As the AMP level is much higher than the ATP level in spores, AMP may be a better biomarker for spore detection. In an AMP-based bioluminescence assay, AMP is converted to ATP using pyruvate orthophosphate dikinase, and ATP is subsequently detected by luciferase. In this present study, we measured the AMP content of several spores of *Bacillus* species using an AMP bioluminescence assay and optimized conditions suitable for rapid spore detection. Spores of several *Bacillus* type strains as well as spacecraft-associated environmental isolates of the same species were purified and compared for AMP content. For the *Bacillus* strains tested, the mean AMP content of a bacterial spore is 10^{17} moles per colony-forming units. Several environmental samples were collected from the spacecraft assembly facility and the sensitivity of the advanced rapid spore detection method was evaluated. The ability of using RSD method to distinguish live vs dead spores was also investigated. In summary, this RSD spore assay is rapid and sensitive and detects spores in real time. The instrument required for the analysis is portable, which makes the RSD method suitable for field deployment and can be integrated. Once integrated this RSD spore detection method may have wide application in homeland security, defense against bio-warfare agents, environmental monitoring, as well as in the food and pharmaceutical industries.

Introduction

Current Spore Detection Methods

Methods	Sensitivity	Speed	Simplicity
Culture	Very High	Very slow	Complicated
Tb-DPA	Low	Rapid	Simple
PCR	High	Rapid	Complicated
Biochip	Low	Rapid	Complicated

Objective: To develop a new sensitive, rapid, and simple spore bioburden detection method.

A Rapid Method for Bio-cleanliness Monitoring

Enzyme Based ATP Assay



Advantages: Very simple, Very rapid, and Sensitive
Disadvantage: Does not detect spores.

AMP Level is much higher than ATP Level in Spores

Adenine Nucleotide Levels in Spores of Various Species

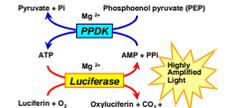
	ATP	ADP	AMP
<i>B. cereus</i>	0.005	0.2	1.3
<i>B. megaterium</i>	0.005	0.2	1.2
<i>B. subtilis</i>	< 0.005	0.2	1.2
<i>C. bifementans</i>	0.006	0.6	2.3
<i>C. perfringens</i>	< 0.05	0.2	1.7
<i>S. halophila</i>	< 0.005	0.1	0.8
<i>S. ureae</i>	< 0.005	0.05	0.5

* Values are reported in micromoles per gram of dry spores. (Lobson, C. A. and Setlow, P. 1993)

AMP may be a better biomarker for spore detection.

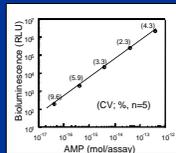
AMP Based Spore Detection

Principle of Bioluminescent Cycling Reaction Using PPKK and Luciferase



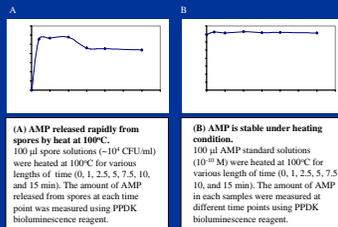
Advantages: Very rapid, more sensitive, CAN detect spores.

Typical Standard Curve for AMP Using PPKK Bioluminescent Reagent

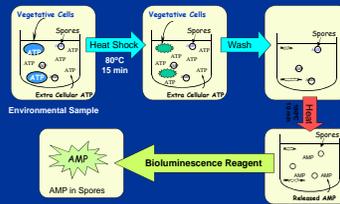


Sensitivity: AMP Detection limit is 10^{17} mol/assay over four magnitudes
Linear Range: coefficient of variation less than 10%
Reproducibility:

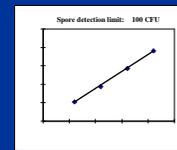
Rapid Release of AMP from Spores by Heat



Procedure for AMP Spore Detection Assay



Sensitivity of AMP Spore Detection Method



100 µl spore solutions (10^7 – 10^8 CFU/ml) were heated at 100°C for 10 min. The amount of AMP released from spores in each sample was measured using PPKK bioluminescence reagent.

Preparation of wild type stain spores

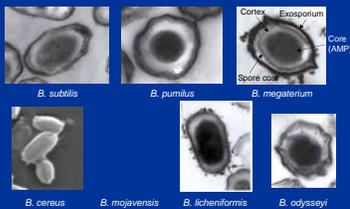
Sample collection:

Wild type strain spores were collected from various classroom facilities from year 2000 to 2002. The isolates were classified using Biolog system (Biolog, Hayward, CA) and identified by 16S ribosomal DNA sequencing.

Spore isolation and purification:

Overnight culture was inoculated on NSM agar-plates and grown at 37°C for two days. The plate was placed at room temperature for an additional day before purification procedure. Spores were purified using water purification method.

Spore Images of Various Bacillus Species

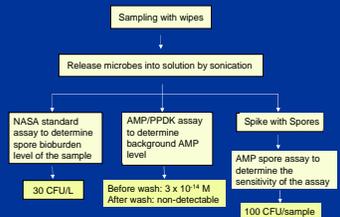


AMP Levels in Various Strains of Spores

Species	Strain	Type Strain	Moles AMP	CFU/assay	AMP/CFU
<i>B. subtilis</i>	ATCC 6051	Type Strain	1.71E-15	220	8.03E-18
	ATCC 168-2	Type Strain	2.82E-16	290	9.74E-19
<i>B. pumilus</i>	FO 340	Wild Type	3.20E-16	140	2.29E-18
<i>B. cereus</i>	FO-11	Wild Type	8.00E-16	96	8.34E-18
<i>B. megaterium</i>	ATCC 14581	Type Strain	1.46E-15	830	1.73E-18
<i>B. megaventsis</i>	ATCC51516	Type Strain	3.34E-16	80	4.18E-18
	KL-154	Wild Type	2.07E-16	450	4.62E-19
<i>B. odyssaei</i>	20411	Wild Type	2.86E-16	200	1.43E-18
<i>B. licheniformis</i>	ATCC14580	Type Strain	1.72E-16	68	2.55E-18
	KL-196	Wild Type	1.75E-15	840	2.08E-18
		Mean			3.21E-18
		STDEV			2.89E-18

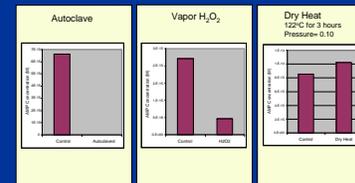
Spores of several *Bacillus* type strains as well as spacecraft-associated environmental isolates of the same species were purified and compared for AMP content. AMP content in a sample was determined from a standard curve that was established with purified chemical AMP. For the *Bacillus* strains tested, the mean AMP content of a bacteria spore is 10^{17} moles per colony forming unit.

Environmental sample



Live vs Dead

Spores were killed by three procedures: autoclave, vapor H₂O₂, and dry heat. AMP levels in dead spores and control live spores were compared.



Summary

This method is very rapid and sensitive.

Detection time: 10 seconds
Assay time: 30 min
Sensitivity (for AMP): 10^{17} moles/assay
Sensitivity (for Spores): 100 CFU

Rapid release of spore biomarker by heat using PCR machine enabled the HTP (high-through-put) processing of large amount of samples in 96 well format.

The instrument required for the analysis is portable, which makes this method suitable for field-applications.

This method can be used to evaluate the sterilization quality of autoclave and H₂O₂ procedure.